A duplicate copy of this sheet is enclosed.

c. ■ The Commissioner is hereby authorized to charge any additional fees which may be required, or credit any overpayment to Deposit Account No. 13-3405. A duplicate copy of this sheet is enclosed.

FORM PTO-1390 (REV. 6-87)

U.S. DEPARTMENT OF COMMERCE PATENT AND TRADEMARK OFFICE.

ATTORNEY'S DOCKET NUMBER

9 9 CFP 1998

29 SEP 1998

# TRANSMITTAL LETTER TO THE UNITED STATES DESIGNATED/ELECTED OFFICE (DO/EO/US)

09/155514

		INTERNATIONAL FILIN 29 January 1998 (29.01.9		PRIORITY DATE CLAIMED 29 January 1997 (29.01.97)		
TITLE OF INVENTION CHIMERIC PROTEINS, THEIR HETERODIMER COMPLEXES, AND PLATELET SUBSTITUTES						
APPLICANT(S) FOR DO/EO/US Mie Kainoh and Toshiaki Tanaka						
Applicant herewith submits to the United States Designated/Elected Office (DO/EO/US) the following items under 35 U.S.C. 371:						
1. ■ This express request to immediately begin national examination procedures (35 U.S.C. 371(f)).						
2. ■ The U.S. National Fee (35 U.S.C. 371(c)(1)) and other fees as follows:						
CLAIMS	(1) FOR	(2) NUMBER FILED	(3) NUMBER EXTRA	(4) RATE	(5) CALCULATIONS	
	TOTAL CLAIMS	64 -20=	44	x \$22.00	\$ 968.00	
	INDEPENDENT CLAIMS	3 -3=	0	x \$82.00		
	MULTIPLE DEPENDENT (	CLAIM(S) (if applicable)		+ \$270.00	270.00	
	BASIC NATIONAL FEE (37 CFR 1.492(a)(1)-(4)):  International preliminary examination fee paid to USPTO (37 CFR 1.482)				,	
					930.00	
			TOTAL	OF ABOVE CALCULATIONS	=2,168.00	
	Reduction by 1/2 for filing by small entity, if applicable. Affidavits must be filed also. (Note 37 CFR 1.9, 1.27, 1.28.)					
				SUBTOTAL	+2,168.00	
	Processing fee of \$ for furnishing the English Translation later than \[ \sum 20 \] \[ \sum 30 \] mos. from the earliest claimed priority date (37 CFR 1.482(f)). \[ \$130.00					
			<u> </u>	TOTAL NATIONAL FEE	\$2,168.00	
	Fee for recording the enclosed assignment (37 CFR 1.21(h)). \$40.00				+	
				TOTAL FEES ENCLOSED	\$2,168.00	
a.  A check in the amount of \$2,168.00 to cover the above fees is enclosed.						
b.   Please charge my Deposit Account No. 13-3405 in the amount of \$ to cover the above fees.						

<ul> <li>3. A copy of the International Application as filed (35 U.S.C. 371(c)(2))</li> <li>a. □ is transmitted herewith (required only if not transmitted by the International Bureau).</li> <li>b. □ is not required, as the application was filed in the United States Receiving Office (RO/US).</li> <li>c. ■ has been transmitted by the International Bureau.</li> </ul>
4. A translation of the International Application into English (35 U.S.C. 371(c)(2)).
<ul> <li>5. Amendments to the claims of the International Application under PCT Article 19 (35 U.S.C. 371(c)(3))</li> <li>a. □ are transmitted herewith (required only if not transmitted by the International Bureau).</li> <li>b. □ have been transmitted by the International Bureau.</li> </ul>
6.   A translation of the amendments to the claims under PCT Article 19 (35 U.S.C. 371(c)(3)).
7.   An oath or declaration of the inventor (35 U.S.C. 371(c)(4)).
8.   A translation of the Annexes to the International Preliminary Examination Report under PCT Article 36 (35 U.S.C. 371(c)(5)).
Other document(s) or information included:
9.   An Information Disclosure Statement under 37 C.F.R. 1.97 and 1.98.
10.   An Assignment document for recording and a Recordation Form Cover Sheet - Patents Only. Please mail the recorded assignment document to the person whose signature, name and address appears at the bottom of this page.
<ul> <li>11. The above checked items are being transmitted</li> <li>a. □ before the 18th month publication.</li> <li>b. ■ after publication and the Article 20 communication but before 20 months from the priority date.</li> <li>c. □ after 20 months but before 22 months (surcharge and/or processing fee included).</li> <li>d. □ after 22 months (surcharge and/or processing fee included).</li> <li>Note: Petition to revive (37 C.F.R. 1.137(a) or (b)) is necessary if 35 U.S.C. 371 requirements submitted after 22 months and no proper demand for International Preliminary Examination was made by 19 months from the earliest claimed priority date.</li> <li>e. □ by 30 months and a proper demand for International Preliminary Examination was made by the 19th month from the earliest claimed priority date.</li> <li>f. □ after 30 months but before 32 months and a proper demand for International Preliminary Examination was made by the 19th month from the earliest claimed priority date (surcharge and/or processing fee included).</li> <li>g. □ after 32 months (surcharge and/or processing fee included).</li> <li>Note: Petition to revive (37 C.F.R. 1.137(a) or (b)) is necessary if 35 U.S.C. 371 requirements submitted after 32 months and a proper demand for International Preliminary Examination was made by 19 months from the earliest claimed priority date.</li> <li>12. At the time of transmittal, the time limit for amending claims under Article 19</li> </ul>
<ul> <li>a. □ has expired and no amendments were made.</li> <li>b. □ has not yet expired.</li> </ul>
13.   Certain requirements under 35 U.S.C. 371 were previously submitted by the applicant on, namely:
Date: 29 SEPT 1993  By: July 31750  Austin R. Miller, Reg. No. 16,602  36th Floor 1600 Market Street Philadelphia, PA 19103 (215) 563-1810

3/PRTS
Express Mail Label EL139842061US

09/155514 405Rec'd PCT/PTO 28 SEP 1998 21 29 SEP 1998

### **SPECIFICATION**

Chimeric proteins, their heterodimer complexes, and platelet substitutes

Technical Field:

The present invention relates to chimeric proteins consisting of an integrin and an immunoglobulin, their heterodimer complexes, a production process thereof, their applications as drugs and reagents, etc. Furthermore, the present invention relates to medicinal application of isolated extracellular maxtrix receptors such as integrin—immunoglobulin chimeric protein heterodimer complexes, as platelet substitutes.

## Background Arts:

Various cells have receptors which mediate the adhesion between a cell and a cell and receptors which mediate the adhesion between a cell and an extracellular matrix, and these receptors play important roles in immune reaction, inflammatory reaction, development, morphogenesis, wound healing, hemostasis, cancerous metastasis, etc. By separating and identifying the receptors which participate in these phenomena, the existence of so-called cell adhesion molecules has been clarified. Many of the molecules identified one after another are classified in reference to their structural features into integrin superfamily, immunoglobulin superfamily, selectin family, cadherin family, etc. (Corlos,

T. M. and Harlan, J. M., Blood, 84, 2068-2101 (1994)). these families, the immunoglobulin superfamily, selectin family and cadherin family mediate mainly the adhesion between a cell and a cell, while the integrin superfamily is the socalled extracellular matrix receptors which mediate the adhesion to extracellular matrices such as fibronectin and collagens. In addition, extracellular matrix receptors which do not belong to any of these adhesion molecule families include CD26 (DDPIV), CD44, GPIV, GPVI, GPIb-vWF, etc. is a receptor for collagens, and CD44 is a receptor for hyaluronic acid, fibronectin and collagens ("Adhesion Molecules" p. 32-42, Masayuki Miyasaka (1991), Medical View (in Japanese)). Furthermore, it is reported that among the membrane glycoproteins (GPs) existing on platelets, GPIV, GPVI, GPIb-vWF, etc. are also collagen receptors ("Platelet Receptors", p. 119-132, Minoru Ohkuma et al., (1992), Kinpodo (in Japanese)).

A receptor belonging to the integrin superfamily has a heterodimer complex structure in which two subunits,  $\alpha$ -chain and  $\beta$ -chain as mutually different membrane proteins are associated with each other non-covalently (Hynes, R. O., Cell, 48, 549-554 (1987)). In the past, the integrin superfamily was classified into three subfamilies;  $\beta$ 1 integrin,  $\beta$ 2 integrin and  $\beta$ 3 integrin. Later, new  $\beta$  chains and  $\alpha$  chains were discovered one after another, and presently eight  $\beta$ 

chains ( $\beta$ 1,  $\beta$ 2  $\beta$ 3,  $\beta$ 4,  $\beta$ 5,  $\beta$ 6,  $\beta$ 7 and  $\beta$ 8 and fifteen  $\alpha$  chains ( $\alpha$ 1,  $\alpha$ 2,  $\alpha$ 3,  $\alpha$ 4,  $\alpha$ 5,  $\alpha$ 6,  $\alpha$ 7,  $\alpha$ 8,  $\alpha$ 9,  $\alpha$ v,  $\alpha$ L,  $\alpha$ M,  $\alpha$ X,  $\alpha$ IIb and  $\alpha$ E) have been identified (Elner, S. G. and Elner, V. M. Inv. Ophtal. Vis. Sci., 37, 696-701 (1996)). It is known that each  $\beta$  chain is associated with one to eight  $\alpha$  chains, and as a result, 21 pairs of an  $\alpha$  chain and a  $\beta$  chain, i.e., integrin molecules have been identified (Elner, S. G. and Elner, V. M., Inv. Ophtal, Vis. Sci., 37, 696-701 (1996)). They include  $\alpha$ 4 $\beta$ 1 (VLA-4,  $\beta$ 1 integrin),  $\alpha$ L $\beta$ 2 (LFA-1,  $\beta$ 2 integrin),  $\alpha$ M $\beta$ 2 (Mac-1,  $\beta$ 2 integrin),  $\alpha$ IIb $\beta$ 3 (GPIIb/IIIa,  $\beta$ 3 integrin), etc. now targeted for drug development (Drug and Market Development, 6, 201-205 (1995)). Many other integrins are also expected to have relations with diseases.

The heterodimer complex structure of an integrin plays an important role in binding to a ligand (Hynes, R. O., Cell, 48, 549-554 (1987)). For example, it is estimated that the ligand binding region on an integrin consists of both an  $\alpha$  chain and a  $\beta$  chain (Hynes, R. O., Cell, 69, 11-25 (1992)). The fact that integrins having the same  $\alpha$  chain but associated with a different  $\beta$  chain, or integrins having the same  $\beta$  chain but associated with a different  $\alpha$  chain are respectively different in substrate specificity (Elner, S. G. and Elner, V. M., Inv. Ophtal, Vis. Sci. 37, 696-701 (1996)) supports this assumption. On the other hand, it was reported that the  $\alpha$  chains of some integrins have an sequence called an I domain

consisting of about 180 amino acids inserted in the molecule, and data suggest that the I domain only could be bound to a ligand were reported (Ueda, T. et al., Proc. Natl. Acad. Sci. USA, 91, 10680-10684 (1994)). However, it was also reported that the I domain of an  $\alpha$  domain and the integrin as its original heterodimer complex are different in the style of binding to a ligand (Kamata, T. and Takada, Y., J. Biol. Chem., 269, 26006-26010 (1994)). It is also not clarified yet whether such parameters as specificity and affinity to a ligand are identical. It is not reported that in the case of an integrin not containing the I domain, for example, in the case of  $\alpha 4\beta 1$  a partial structure only is bound to a ligand.

If any integrin isolated and prepared retains its heterodimer complex structure, hence the ligand binding capability, it can be used for studying the style of binding to a ligand in a state close to nature. Furthermore, it can be used as it is as a drug and can also be used as a reagent for measuring the amount of a ligand in tissue or serum or as a material for searching for adhesion inhibiting compounds very usefully. However, isolating and preparing an integrin with its function retained is said to be very difficult. One reason is that since the association between an  $\alpha$  chain and a  $\beta$  chain of an integrin is maintained non-covalently as desribed before, they are easily dissociated during isolation and preparation. Since an integrin is a membrane protein, the

necessity of using a surfactant, etc. for solubilization is considered to be a large cause in the dissociation of the complex. In other words, the non-covalent preservation of functional structure inhibits the preparation of such an integrin.

Inspite of the difficulty as described above, some cases were reported, in which an integrin heterodimer complex was isolated and prepared with its function retained. For cases of  $\alpha 2\beta 1$ ,  $\alpha 5\beta 1$  and  $\alpha \vee \beta 3$ , it was reported that the binding to a ligand can be determined by letting a liposome incorporate an integrin purified by using affinity column chromatography (Santoro, S. A. et al., Biochem. Biophys. Res. Comm., 153, 217-223 (1988), Pytela, R. et al. Cell, 40, 191-198 (1985), Pytela, R. et al., Method Enzymol., 144, 475-489 (1987)). For other cases, it was that if purified  $\alpha 5 \beta 1$  or  $\alpha v \beta 3$  is coated on a plate, a peptide which inhibits the cell adhesion through the integrin can be selected (Koivunen, E. et al., J. Biol. Chem., 268, 20205-20210 (1993), Healy, J. M. et al., Biochemistry, 34, 3948-3955 (1995)). For further other cases, it was reported that if purified  $\alpha \vee \beta 3$  or  $\alpha 4 \beta 1$  is coated on a plate, the binding to a ligand can be determined (Charo, I. F. et al., J. Cell Biol., III, 2795-2800 (1990), Makarem, R. et al., J. Biol. Chem., 269, 4005-4011 (1994), Paul Mould, A. et al., J. Biol. Chem., 269, 27224-27230 (1994)). For a still further other case, it was reported that if an extracellar

portion of  $\alpha$ Ilb $\beta$ 3 heterodimer complex prepared by gene manipulation is coated on a plate through a complex specific antibody, the binding to a ligand can be determined (Gulino, D. et al., Eur. J. Biochem., 227, 108-115 (1995)). These cases suggest that to exert the function of a purified integrin, its heterodimer complex must be bound to or included in any carrier. The reason why a carrier is considered to be necessary is that since a heterodimeter complex is associated non-covalently in a solution, it tends to be dissociated and as a result, cannot retain its fuctional structure. In the finally stated case, only a molecule with a heterodimer complex structure is selected using a complex specific antibody, in a design to determine the binding in a state where both the chains are not dissociated from each other.

As a case requiring no carrier, it was reported that purified  $\alpha 1 \beta 1$ , or  $\alpha 2 \beta 1$  allows the determination of the bonding to a ligand dependent on high concentration of metal ions even without using any carrier (Pfaff, M. et al., Eur. J. Biochem., 225, 975-984 (1994)). In this case, the surfactant added in the process of purification plays a role similar to that of a liposome, acting as a carrier. For a further other case, it was reported that an extracellular of  $\alpha M \beta 2$  heterodimer complex prepared by using gene manipulation is bound to a ligand (Berman, P. W. et al., J. Cell Biochem., 52, 183-195 (1993)). These cases do not suggest the necessity of

any carrier as described before, but the disadvantage that the association of molecules in a heterodimer complex is retained non-covalently is not improved.

As a still further other case, a chimeric protein consisting of  $\alpha d$  and an immunoglobulin is disclosed (Japanese Patent Laid-Open (Kokai) No. 8-507933), but only the result of immune precipitation is reported, without examining the binding to a ligand. Furthermore, since a  $\beta$  chain is not expressed in the chimeric protein as an immunoglobulin, the binding between an  $\alpha$  chain and a  $\beta$  chain remains non-covalent.

The above facts suggest that any integrin with an  $\alpha$  chain and a  $\beta$  chain structurally stably associated and with its function retained has never been successfully prepared. That a complex structure is unstable restricts the use of its molecule.

Of the molecules belonging to the integrin superfamily, integrin  $\alpha 2\beta 1$  is an extracellular matrix receptor found to be expressed in T cells, platelets, etc. activated for long time. However, it was reported that the  $\alpha 2\beta 1$  on the cell surfaces of platelets and fibroblasts is bound to collagens only and that the  $\alpha 2\beta 1$  on the surfaces of vascular endothelial cells is bound to both collagens and laminins (Elices, M. J. et al., Proc. Natl. Acad. Sci. USA, 86, 9906-9910 (1989)), and it is speculated that the function of  $\alpha 2\beta 1$  becomes different,

depending on cells.

In relation to the conditions of diseases, there are reports to suggest that integrin  $\alpha 2\beta 1$  plays an important roll for wound healing and cancerous metastasis (Shiro, J. A. et al., Cell, 67, 403-410 (1991), Chen, F. et al., J. Exp. Med., 173, 1111-1119 (1991), Chan, B. M. C. et al., Science, 251, 1600-1602 (1991)). Furthermore, it was reported that from the analysis of platelet function of patients with bleeding tendency, the adherence of platelets and collagens through integrin  $\alpha 2\beta 1$  has close relation with the first step of hemostasis and thrombosis process (Nieuwenhuis, H. K. et al., Nature, 318, 470-472 (1985)). Though the relations of integrin  $\alpha 2\beta 1$  with conditions of diseases are suggested like this, any medical application of using the integrin  $\alpha 2\beta 1$ protein and other isolated extracellular matrix receptor proteins under physiological ion condition or in the presence of plasma components has not been examined.

On the other hand, the necessity for artificial substitutes of platelets used as blood preparations in the clinical field is growing, and various attempts have been reported (Progress of Medicine 179, 406-407 (1996), Clinical Blood 37, 1353-1361 (1997) (respectively in Japanese)). However, they are not yet practically available.

The present invention relates to chimeric proteins in

which the lpha chain and eta chain of an integrin are combined with the heavy chain or light chain of an immunoglobulin, their heterodimer complexes, a production process thereof, a method for testing the binding of an integrin-immunoglobulin chimeric protein heterodimer complex to a ligand and a cell, substances bound to an integrin obtained by using the method, a method for searching for a substance inhibiting the binding between an integrin and a ligand using the integrinimmunogobulin chimeric protein heterodimer complex, substances for inhibiting the binding, and the application of integrinimmunoglobulin chimeric protein heterodimer complexes as drugs and reagents. Furthermore, the present invention relates to platelet substitutes containing an integrin-immunoglobulin chimeric protein heterodimer complex or any other isolated extracllular matrix receptor as an active ingredient. Brief Description of the Drawings:

Fig. 1 shows that  $\alpha 4 \cdot \lg G$  heavy chain- $\beta 1 \cdot \lg G$  heavy chain chimeric protein heterodimer complex is bound to VCAM-1 expressing cell, and that the binding is inhibited by an anti-integrin antibody or EDTA, a cationic chelating agent.

Fig. 2 shows that  $\alpha 4 \cdot \lg G$  heavy chain- $\beta 1 \cdot \lg G$  heavy chain chimeric protein heterodimer complex is bound to CS-1 peptide, and that the binding is inhibited by an anti-integrin antibody or EDTA, a cationic chelating agent.

Fig. 3 shows that the binding between  $\alpha 4 \cdot \lg G$  heavy

chain- $\beta$ 1lgG heavy chain chimeric protein heterodimer complex and CS-1 piptide is inhibited by GPEILDVPST, and is not inhibited by any other peptide.

Fig. 4 shows that  $\alpha 2 \cdot \lg G$  heavy chain— $\beta \lg G$  heavy chain chimeric protein heterodimer complex is bound to a collagen, and that the binding is inhibited by an anti-integrin antibody and EDTA, a cationic chelating agent.

Fig. 5 shows that  $\alpha 2 \cdot \lg G$  heavy chain- $\beta 1 \lg G$  heavy chain chimeric protein heterodimer complex liposome is bound to a collagen in the presence of plasma.

Fig. 6 shows that the binding of  $\alpha 2 \cdot \lg G$  heavy chain- $\beta$  11gG heavy chain chimeric protein heterodimer complex liposome to a collagen is inhibited by an anti-integrin antibody or EDTA, a cationic chelating agent.

The Best Embodiments of the Invention:

The extracellular matrix receptors in the present invention refer generally to the receptors which mediate the adhesion between a cell and an extracellular matrix. The receptors include the integrin superfamily having a heterodimer complex structure in which an  $\alpha$  chain and a  $\beta$  chain are non-covalently associated with each other as two membrane proteins (Corlos, T. M. and Harlan, J. M. Blood, 84, 2068-2101 (1994)), and other receptors such as CD26 (DDPIV), CD44, GPIV, GPVI, GPb-vWF, etc. The integrins in the present invention refer to molecules belonging to the integrin

superfamily, and also include the isomers of the molecules belonging to the family. The  $\alpha$  chains of the present invention include 15  $\alpha$  chains, i.e.,  $\alpha 1, \alpha 2, \alpha 3, \alpha 4, \alpha 5, \alpha 6, \alpha 7, \alpha 8, \alpha 9, \alpha v, \alpha L, \alpha M, \alpha X, \alpha IIb$  and  $\alpha E$ , and among them,  $\alpha 4$  and  $\alpha 2$  are preferable, though preferable  $\alpha$  chains are not limited to them. The  $\beta$  chains of the present invention include eight  $\beta$  chains, i.e.,  $\beta 1, \beta 2, \beta 3, \beta 4, \beta 5, \beta 6, \beta 7$  and  $\beta 8$  and among them,  $\beta 1$  is preferable, though preferable  $\beta$  chains are not limited to it. The integrin molecules as pairs respectively consisting of an  $\alpha$  chain and a  $\beta$  chain include the twenty one integrins stated in Elner, S. G. and Elner, V. M., Inv. Ophtal. Vis. Sci., 37, 696-701 (1996), though not limited to them.

A chimeric protein consisting of the  $\alpha$  chain of an integrin and the heavy chain or light chain of an immunoglobulin refers to a molecule in which the extracellular region of the  $\alpha$  chain of an integrin is bound to the constant region of the heavy chain or light chain contained an immunoglobulin. In this case, a chimeric protein in which N terminus side of the protein is integrin molecule and then connected to an immunoglobulin molecule side by side is preferable. A chimeric protein consisting of the  $\beta$  chain of an integrin and the heavy chain or light chain of an immunoglobulin refers to a molecule in which the extracellular region of the  $\beta$  chain of an integrin is bound to the constant

region of the heavy chain or light chain contained in an immunoglobulin. Also in this case, a chimeric protein in which N terminus side of the protein is an integrin molecule and then connected to an immunoglobulin molecule side by side is preferable. In either case of  $\alpha$  chain or  $\beta$  chain, a chimeric protein bound to the heavy chain of an immunoglobulin is preferable.

The isotype of the immunoglobulin to be bound to the  $\alpha$  chain or  $\beta$  chain is not especially limited. Any of lgG, lgM, lgA and lgE can be used, but it is preferable to use lgG. The subclasses of lgG include lgG1, lgG2, lgG3 and lgG4, but it is preferable to use lgG1. Furthermore, it is possible to use a molecule with a dimer structure having a disulfide bond between molecules instead of the immunoglobulin.

In the present invention, a molecule in which a chimeric protein consisting of the  $\alpha$  chain of an integrin and the heavy chain or light chain of an immunoglobulin and a chimeric protein consisting of the  $\beta$  chain of the integrin and the havy chain or light chain of the immunoglobuline are associated with each other is called an integrin-immunoglobulin chimeric protein heterodimer complex. In this case, a combination consisting of  $\alpha$  chain-immunoglobulin heavy chain (which means a chimeric protein consisting of an  $\alpha$  chain and the heavy chain of an immunoglobulin; hereinafter this applies) and  $\beta$  chain-immunoglobulin heavy chain, a combination consisting of

 $\alpha$  chain immunoglobulin heavy chain and  $\beta$  chain immunoglobulin light chain, and a combination consisting of  $\alpha$  chain immunoglobulin light chain and  $\beta$  chain immunoglobulin heavy chain are preferable. A combination consisting of  $\alpha$  chain immunoglobulin heavy chain and  $\beta$  chain immunoglobulin heavy chain is more preferable.

In the integrin-immunoglobulin chimeric protein heterodimer complex of the present invention, the  $\alpha$  chain can be  $\alpha 1$ ,  $\alpha 2$ ,  $\alpha 3$ ,  $\alpha 4$ ,  $\alpha 5$ ,  $\alpha 6$ ,  $\alpha 7$ ,  $\alpha 8$ ,  $\alpha 9$ ,  $\alpha v$ ,  $\alpha L$ ,  $\alpha M$ ,  $\alpha X$ ,  $\alpha IIb$  or  $\alpha E$ , and the  $\beta$  chain can be  $\beta 1$ ,  $\beta 2$ ,  $\beta 3$ ,  $\beta 4$ ,  $\beta 5$ ,  $\beta 6$ ,  $\beta 7$  or  $\beta 8$ , It is preferable that the  $\alpha$  chain is  $\alpha 4$  or  $\alpha 2$  and that the  $\beta$  chain is  $\beta 1$ , though preferable chains are not limited to them.

The process for preparing an integrin-immunogloburin chimeric protein heterodimer complex is described below, but the process is not limited thereto.

A DNA coding for the  $\alpha$  chain and  $\beta$  chain of an integrin can be obtained using the information of known cDNA sequences by such a method as gene amplification based on the PCR method, cDNA cloning or DNA synthesis. For example, the DNA sequences of  $\alpha$ 4 and  $\beta$ 1 are already reported in literature (Takada, Y. et al., EMBO J., 8, 1361-1368 (1989), Scott Argraves, W. et al., J. Cell Biol., 105, 1183-1190 (1987)). A DNA coding for the  $\alpha$  chain and  $\beta$  chain of an integrin can also be obtained by the expression cloning using an antibody, etc. For binding to a DNA coding for the constant region of

an immunoglobulin, it is desirable to take out a DNA coding for the extracellular portions only of the lpha chain and etachain of an integrin. For this purpose, it is preferable to use the PCR method and DNA synthesis. The extracellular portion of either an  $\alpha$  chain or  $\beta$  chain refers to the polypeptide sequence on the N terminus side from the portion speculated to be the transmembrane portion. Its partial sequence can also be used as far as the ligand binding capability is retained, but it is preferable to use most of the portion considered to be an extracellular region. For taking out a DNA, it is necessary to adjust for adaptation of frames after linking to a DNA coding for an immunoglobulin. For example, this can be achieved by modifying the primer when a DNA fragment is taken out by the PCR method. In this case, it is desirable to design for ensuring that amino acid modification is not caused by the base substitution of the primer. However, amino acid substitution is allowed as far as the function of the chimeric protein is not changed. For obtaining a DNA by chemical synthesis, the purpose can be achieved by designing a sequence to ensure the linking to a DNA coding for an immunoglobulin. In the case of cDNA, a DNA capable of being bound to a DNA coding for an immunoglobulin can be prepared by using DNA fragmentation and a synthetic DNA.

Then, a DNA coding for an immunoglobulin is prepared. In

the present invention, it is desirable to use DNAs coding for the heavy chain and light chain of a human immunoglobulin, but DNAs coding for an immunoglobulin of another animal species can also be used. The preparation of a DNA coding for human lgG is already reported (Ellison, J. W. et al., Nucleic Acids Res., 10, 4071-4079 (1982)), but the preparation is not limited to this method. Any method similar to the above mentioned method for preparing DNAs coding for the lpha chain and B chain of an integrin can also be used. In the present invention, for the heavy chain of a human immunoglobulin, it is preferable to use a genomio DNA, but a cDNA can also be used. As the DNA for the heavy chain of a human immunoglobulin, it is preferable to use a portion coding for the hinge region, CH2 region or CH3 region, but a DNA coding for the entire constant region of CH1 — CH3 can also be used. For the light chain of an immunoglobulin, a DNA coding for the CL region is used. Finally, a DNA coding for the extracellular portion of an  $\alpha$  chain or  $\beta$  chain and a DNA coding for the constant region of human immunoglobulin heavy chain are linked with in frame. The obtained DNA codes for a polypeptide starting from the methionine of translation initiation and having the signal sequence of the lpha chain or etachain of an integrin, its extracellular region and the constant region of human immunoglobulin heavy chain linked in this order.

The DNA coding for a chimeric protein consisting of the lphachain of an integrin and the heavy chain or light chain of an immunoglobulin, or the DNA coding for a chimeric protein consisting of the  $oldsymbol{eta}$  chain of an integrin and the heavy chain or light chain of an immunoglobulin respectively obtained in the above is functionally linked in a proper expression control sequence, to obtain a recombinant vector. The general methods concerning gene recombination such as the method for preparing the recombinant vector, the method for transfecting it into a cell are described in a published book ("Molecular Cloning", Sambrook et al., (1989) Cold Spring Harbor Lab. Press, New York), but the methods are not limited to those stated there. In the present invention, it is desirable to use an expression control sequence suitable for protein expression in animal cells. For example, for manifestation of insect cells, polyhedrin promotor, p10 promotor, etc. are generally used as expression control sequences, and for expression of other animals' cells,  $SR\alpha$  promotor, cytomegalovirus derived promotor, simian virus 40 derived promotor, polyhedrin promotor, p10 promotor, etc. are used. However, the expression control sequences are not limited to them. In the present invention, it is preferable to use SRlphapromotor.

If the obtained recombinant vector is transfected into a cell, a cell capable of producing an integrin-immunoglobulin

chimeric protein heterodimer complex can be obtained. In this case it is preferable to use an animal derived cell as a host. For example, COS cell (simian renal cell), CHO cell (Chinese Hamster ovarian cell), Sf9 (insect cell), etc. are generally used as hosts. Furthermore, myeloma cells such as P3U1 and Y3 can also be used. Other established cell lines and cloned cells can also be used, but the cells used as hosts are not limited to them. In the present invention, it is preferable to use a CHO cell.

It is known that the methods for transfecting a recombinant vector into a cell include the lipofectin method, calcium phosphate method, electroporation method, etc., and any of the methods can be used. The method is not limited to them. It is preferable that when a cell is transfected by using a recombinant vector, a recombinant vector for expression of a chimeric protein consisting of the  $\alpha$  chain of an integrin and the heavy chain or light chain of an immunoglobulin and a recombinant vector for expression of a chimeric protein consisting of the eta chain of the integrin and the heavy chain or light chain of the immunoglobulin are transfected into the cell one after another using different drug resistance markers. The recombinant vectors can be transfected in any order or simultaneously. It is desirable that the two recombinant vectors to be transfected are vectors for expression of a combination consisting of lpha chain.

immunoglobulin heavy chain (which means a chimeric protein consisting of an  $\alpha$  chain and the heavy chain of an immunoglobulin; hereinafter this applies) and  $\beta$  chain immunoglobulin heavy chain, or  $\alpha$  chain immunoglobulin heavy chain and  $\beta$  chain immunoglobulin light chain, or  $\alpha$  chain immunoglobulin light chain and  $\beta$  chain immunoglobulin heavy chain. Any of these combinations can be adopted, but a combination of recombinant vectors for expression of  $\alpha$  chain immunoglobulin heavy chain and  $\beta$  chain immunoglobulin heavy chain is desirable.

In any transfection method and any combination of vectors, it is important to select a cell which is transfected by the two recombinant vectors and produces a chimera protein consisting of an  $\alpha$  chain and the heavy chain or light chain of an immunoglobulin and a chimera protein consisting of a  $\beta$  chain and the heavy chain or light chain of an immunoglobulin simultaneously almost by the same amounts. This can be achieved by measuring the amounts of the chimeric protein consisting of an  $\alpha$  chain and the heavy chain or light chain of an immunoglobulin and a chimeric protein consisting of a  $\beta$  chain and the heavy chain or light chain of an immunoglobulin produced in the cultured supernatant solution of the cell transfected by the recombinant vectors. For measurement, for example, the transfected cell can be cultured in a medium containing  $^{35}$ S according to any publicly known method, for

labeling the proteins, and the amounts of the chimeric protein consisting of an  $\alpha$  chain and the heavy chain or light chain of an immunoglobulin and a chimeric protein consisting of a  $oldsymbol{eta}$ chain and the heavy chain or light chain of an immunoglobulin existing in the cultured supernatant solution can be estimated by immunoprecipitation using an anti- $\alpha$  chain antibody or an anti- $\beta$  chain antibody respectively. As another method, the amounts the chimeric protein consisting of an lpha chain and the heavy chain or light chain of an immunoglobulin and a chimeric protein consisting of a  $\beta$  chain and the heavy chain or light chain of an immunoglobulin existing in the cultured supernatant solution can be estimated according to the ELISA method using an anti-human immunoglobulin antibody and an anti- $\alpha$  chain antibody or an anti- $\beta$  chain antibody. Anyway, it is preferable to select a clone which produces almost the same large amounts of the chimeric proteins of the lpha and etachains in the culture supernatant , for preparing an integrinimmunoglobulin chimeric protein heterodimer complex. methods for labeling proteins, the methods of immunoprecipitation and the general methods of ELISA are described in a published book ("Antibody" Harlow, E., and Lane, D. (1988), Cold Spring Harbor Lab. Press, New York), but the methods are not limited to them. Any other method can also be used for detecting chimeric proteins.

The obtained transfected cell can be cultured according to

a general cell culture method, to produce an integrinimmunoglobulin chimeric protein heterodimer complex. It is
preferable that the medium contains about 5% of serum of a low
immunoglobulin concentration, but any generally known serumcontaining medium or a serum-less medium can also be used.
After completion of cell culture, the cells and solid matter
are removed by such operation as centrifugation, and the
culture supernatant containing an integrin-immunoglobulin
chimeric protein heterodimer complex is collected.

It can be estimated that the cultured supernatant solution contains not only the integrin-immunoglobulin chimeric proteins in which the chimeric protein consisting of an lphachain and the heavy chain or light chain of an immunoglobulin and a chimeric protein consisting of a  $oldsymbol{eta}$  chain and the heavy chain or light chain of an immunoglobulin form a heterodimer complex, but also the chimeric protein consisting of an lphachain and the heavy chain or light chain of an immunoglobulin and a chimeric protein consisting of a eta chain and the heavy chain or light chain of an immunoglobulin which do not form the heterodimer complex. However, since the molecules other than the heterodimer complex cannot be bound to a ligand, the supernatant solution can be used as a reagent for testing the binding to a ligand or cell, or searching for a substance inhibiting the binding between an integrin and a ligand, or for searching for a substance capable of being bound to an

integrin, or for measuring the ligand amount of an integrin.

These methods of utilization are basically the same as those for using a purified integrin-immunoglobulin chimeric protein heterodimer complex described later.

An integrin-immunoglobulin chimeric protein heterodimer complex can be purified by an established method using a protein A column chromatography by use of the nature of the immunoglobulin portion. Furthermore, affinity chromatography using an antibody against the  $\alpha$  or  $\beta$  chain can also be used. Moreover, the purification can also be achieved by affinity chromatography with a ligand bound to a carrier. General chromatographic methods can also be used in combination for the purification. If publicly known cases in which integrin molecules are purified by these methods (Pytela, R. et al., Methods Enzymol., 144, 475-489 (1987), Santoro, S. A. et al., Biochem. Biophys. Res. Comm., 153, 217-223 (1988), Charo, I.F. et al., J. Cell Biol., 111, 2795-2800 (1990), Makarem, R. et al., J. Biol. Chem., 269, 4005-4011 (1994), Pfaff, M. et al., Eur. J. Immunol., 225, 975-984 (1994), Gulino, D. et al., Eur. J. Biochem., 227, 108-115 (1995), etc.) are applied, the purification of an integrin-immunoglobulin chimeric protein heterodimer complex can be achieved.

A purified integrin-immunoglobulin chimeric protein heterodimer complex can be identified as a protein showing at least one band under non-reducing condition and at least two

bands under reducting condition by SDS-PAGE. It can also be confirmed from it, that the heterodimer is linked by the disulfide bond between immunoglobulin heavy chains. It sometimes occurs that a plurality of bands are detected under reduction, but this is considered to be probably because intramolecular clearage of the  $\alpha$  chain has occurred. Especially, with lpha4,this phenomenon is known (Hemler, M.E. et al., J. Biol. Chem., 262, 11478-11485 (1987)). Furthermore, it can be confirmed by the Western blotting method that the respective bands indicate chimeric proteins. As another method, it can be confirmed by said ELISA method combining an anti- $\alpha$  chain antibody, anti- $\beta$  chain antibody and anti-human immunoglobulin antibody, that the obtained molecule is an integrin-immunoglobulin chimeric protein heterodimer complex. That is, the molecule can be identified as a protein molecule with epitopes for all the antibodies. As a further other method, an integrin-immunoglobulin chimeric protein heterodimer complex can also be identified by immunoprecipitation. In this case, if the purified protein is labeled by  $^{35}$ S, or  $^{125}$ I or biotin, etc. according to any known method, and immunoprecipitated using an anti- lpha chain antibody, anti- $oldsymbol{eta}$  chain antibody and anti-human immunoglobulin antibody, the same electrophoretic pattern can be obtained in every So, it can be confirmed that the integrincase. immunoglobulin chimeric protein heterodimer complex has the

intended structure. Furthermore, even if a condition to dissociate the integrin complex on a cell membrane such as the coexistence of EDTA or boiling in the presence of SDS is applied, the immunoprecipitation pattern is not changed. So, it can be confirmed that the obtained integrin-immunoglobulin chimeric protein heterodimer complex is a structurally stabilized complex. The methods for confirming an integrin-immunoglobulin chimeric protein heterodimer complex are not limited to those stated above.

The binding between a prepared integrin-immunoglobulin chimeric protein heterodimer complex and a ligand can be tested as described below. After a ligand and an integrinimmunoglobulin chimeric protein heterodimer complex are brought into contact with each other, to make a mixture, the amount of the integrin-immunoglobulin chimeric protein heterodimeter complex bound to the ligand or the amount of the ligand bound to the integrin-immunoglobulin chimeric protein heterodimeter complex is measured. The amount of an integrinimmunoglobulin chimeric protein heterodimer complex can be measured by labeling the complex itself by a fluorescent dye or enzyme or radioisotope, etc. The amount of a ligand can also be measured by any similar method. A detection method such as SPA (Amasham) can also be used for the measurement. Furthermore, any reagent which can recognize a complex or ligand labeled by a fluorescent dye, enzyme or radioisotope,

etc. can also be used for the measurement. The reagent for recognizing an integrin-immunoglobulin chimeric protein heterodimer complex can, for example, be an anti-human immunoglobulin antibody. In this test, it is preferable to bind the molecule to be detected, to any carrier such as a bead or plate. As a ligand, its entire molecule can be used, but a portion retaining the binding activity to an integrin can also be taken out for use. For example, for integrin  $\alpha 4\beta$ 1 or integrin  $\alpha 2\beta$ 1, its ligand, fibronectin or collagen or its peptide fragment bound to a carrier can also be used.

Methods similar to the above can be used to test the binding between an integrin-immunoglobulin chimeric protein heterodimer complex and cells. The amount of the cells bound to a complex can be measured by labeling the cells by a fluorescent dye or radioisotope or using a reagent reacting with the cells, for example, an antibody reacting with a surface antigen. If something like a tissue section is used instead of cells, the amount of the bound integrin-immunoglobulin chimeric protein heterodimer complex is measured by any of the above mentioned methods.

The methods for examining the binding between an integrin-immunoglobulin chimeric protein heterodimer complex and a ligand or cell described above can be used for obtaining a substance inhibiting the binding between an integrin and a ligand, for example, for obtaining an antibody, polypeptide,

peptide or low molecular weight compound. It is preferable to premix a sample and an integrin-immunoglobulin chimeric protein heterodimer complex, and then to measure the amount of the integrin-immunoglobulin chimeric protein heterodimer complex bound to a ligand in any of the above mentioned measuring systems. If the amount of the bound integrin-immunoglobulin chimeric protein heterodimer complex is lowered by adding a certain sample, it can be judged that the sample has inhibitory activity. However, in this sytem, a substance with metal ion chelating action or a substance with surfactant action, etc. may give a false positive result. The sources of samples used include the following integrin bound substances, peptide fragments of ligands, their derivatives, marketed compounds, etc., but are not limited to them.

A case where a purified integrin was coated on a plate to search for a peptide to be bound was reported (Healy, J. M. et al., Biochemistry 34, 3948-3955 (1995)). Even if the integrin-immunoglobulin chimeric protein heterodimer complex obtained in the present invention is used, a substance to be bound to an integrin can be similarly searched for. Especially when the chemiric protein heterodimer complex of the present invention is used, the operation to remove the non-specifically bound substances can be effected under more severe conditions. So, the operation can be simplified advantageously. Furthermore, since the complex is not

dissociated during operation, a bound substance can be selected more specifically advantageously. Known sources suitable for selecting bound substances include a phage peptide library (e.g., Scott, J. K. and Smith, G. P., Science, 249, 386-390 (1990)) and a DNA oligomer library (e.g., O'Connel, D. et al., Proc. Natl. Acad. Sci. USA, 93, 5883-5887 (1996), but in the present invention, it is preferable to use the former.

Furthermore, the method of testing the binding between an integrin-immunoglobulin chimeric protein heterodimer complex and a ligand or cell can also be used as a method for measuring the amount of an integrin ligand in a body fluid or tissue.

Moreover, the integrin-immunoglobulin chimeric protein heterodimer complexes of the present invention can also be used as drugs. The present invention has clarified that integrins and other isolated extracellular matrix receptors can be used as platelet substitutes.

An extracellular matrix receptor preferably used as a platelet substitute is an integrin. The  $\alpha$  chain of the integrin can be  $\alpha 1, \alpha 2, \alpha 3, \alpha 4, \alpha 5, \alpha 6, \alpha 7, \alpha 8, \alpha 9, \alpha v, \alpha L, \alpha M, \alpha X, \alpha IIb or <math>\alpha E$ , and among them,  $\alpha 2$  is preferable. The  $\beta$  chain can be  $\beta 1, \beta 2, \beta 3, \beta 4, \beta 5, \beta 6, \beta 7$  or  $\beta 8$ , and among them,  $\beta 1$  is preferable. Integrin  $\alpha 2\beta 1$  is more preferable. The receptor source for isolation can be a tissue or cell expressing an

extracellular matrix receptor, or a dissolved membrane fraction of a receptor expressing cell prepared by gene manipulation, etc. It is more preferable to design for obtaining a soluble protein by modifying a receptor gene by gene recombination, and to use the cultured supernatant solution of the cells capable of producing it, as a source. Furthermore in the design of the soluble protein, it is preferable that the functional structure of the extracellular matrix receptor is retained. For example, it is desirable to use an integrin-immunoglobulin chimeric protein heterodimer complex obtained by modifying the heterodimer structure of an integrin to allow its  $\alpha$  and  $\beta$  chains to be covalently associated with each other. As the integrin-immunoglobulin chimeric protein heterodimer complex, it is preferable that the  $\alpha$  chain of the integrin is  $\alpha 1, \alpha 2, \alpha 3, \alpha 4, \alpha 5, \alpha 6, \alpha 7, \alpha 8, \alpha$ 9,  $\alpha$ v,  $\alpha$ L,  $\alpha$ M,  $\alpha$ X,  $\alpha$ Ilb or  $\alpha$ E, and among them,  $\alpha$ 2 is more preferable. Furthermore, it is preferable that the  $\beta$  chain is  $\beta$ 1,  $\beta$ 2,  $\beta$ 3,  $\beta$ 4,  $\beta$ 5,  $\beta$ 6,  $\beta$ 7 or  $\beta$ 8, and among them,  $\beta$ 1 is more preferable. It is further more preferable that the lpha chain is  $\alpha$ 2 and that the  $\beta$  chain is  $\beta$ 1. The platelet substitute of the present invention is described below mainly in reference to a typical extracellular matrix receptor, integrin  $\alpha 2\beta$ 1-immunoglolublin chimeric protein heterodimer complex, but the present invention is not limited thereto or thereby.

To confirm the applicability of a purified integrin-

immunoglobulin chimeric protein heterodimer complex as a drug, the purified protein itself is used for examining its pharmacological activity. For obtaining higher capability of being bound to an extracellular matrix, it is more preferable to use an integrin-immunoglobulin chimeric protein heterodimer complex bound to a carrier such as a lipid or protein polymer, etc., but the present invention is not limited to this method.

For use as a platelet substitute, it is preferable to bind an integrin  $\alpha 2\beta 1$ -immunoglolublin chimeric protein heterodimer complex to a liposome covalently according to the method stated in a report (Martin, F. J. et al., Biochemistry, 20, 4229 (1981)). The carrier can also be any other drug carrier than a liposome as far as its use for drugs is permitted. If a liposome is used as the carrier, the liposome is prepared according to the composition and method stated in a published book "Preparation and Experiments of Liposomes (in Japanese)", 0ku, N. (1994), Hirokawa Shoten), but a preferable method is such that the epitope bound to the extracellular matrix of an integrin  $\alpha 2\beta 1$ -immunoglolublin chimeric protein heterodimer complex is exposed outside the liposome membrane.

For confirming that an integrin  $\alpha 2\beta 1$ -immunoglolublin chimeric protein heterodimer complex is bound on the prepared liposome carrier, a flow cytometer is used. The reagents which can be used for recognizing the integrin  $\alpha 2\beta$  1-immunoglolublin chimeric protein heterodimer complex include

an anti-integrin  $\alpha 2$  antibody, anti-integrin  $\beta 1$  antibody, anti-human immunoglobulin antibody, etc. If the antibody used is fluorescently labeled, it can be used for determination directly, but if it is not fluorescently labeled, a secondary antibody which recognizes the immunoglobulin class of the animal species used for preparing the antibody is used as a fluorescent label. As a further other confirmation method, the integrin  $\alpha 2\beta 1$ -immunoglolublin chimeric protein heterodimer composite itself can be labeled by an enzyme or radioisotope, etc., for confirmation in proper combination with a color dye or radioactivity measuring instrument, etc.

To examine the extracellular matrix binding capability using an integrin  $\alpha 2\beta 1$ -immunoglolublin chimeric protein heterodimer complex liposome, it is preferable to suspend the integrin  $\alpha 2\beta 1$ -immunoglolublin chimeric protein heterodimer complex liposome into a buffer with a physiological cation concentration or plasma. The buffer with a physiological cation concentration refers to a buffer containing at least cations such as Mg ions or Ca ions and adjusted to about neutrality. The plasma is prepared by processing the blood collected in the presence of an anticoagulant, according to a general plasma preparation method. As the anticoagulant, for example, heparin or EDTA solution can be added by sufficient units. A marketed normal plasma, coagulation factor deficient plasma or serum, etc. can also be used. However, if the

anticoagulant used lowers the cation concentration, cations are added to achieve a physiological concentration later. Then, the integrin  $\alpha 2\beta 1$ -immunoglolublin chimeric protein heterodimer complex liposome is mixed with an extracellular matrix or its fragment coated on a carrier for a certain time, to judge whether binding takes place. It is preferable that the coating of the extracellular matrix or its fragment as a solid phase is achieved by using a plastic plate, etc., but marketed beads for coating an extracellular matrix as a solid phase, etc. can also be used. When a collagen is used as the extracellular matrix, any animal species and type can be used. The binding reaction between an integrin  $\alpha 2\beta 1$ -immunoglolublin chimeric protein heterodimer complex liposome and an extracellular matrix is effected according to a general method adopted for observing the adherence reaction of platelets. many cases, they are allowed to stand mainly in a static system for a certain time, to induce binding to the matrix, but it is preferable to apply a shaking or shear stress, etc.

The integrin  $\alpha 2\beta 1$ -immunoglolublin chimeric protein heterodimer complex liposome is bound to an extracellular matrix under the conditions as described above, and the amount of binding is measured by applying the above mentioned ELISA method using an anti-human immunoglobulin antibody. For more accurate determination, it is desirable to immobilize the liposome bound to the matrix by 1% glutaraldehyde, etc. As

another method than the ELISA method, for example, if a radiolabeled lipid is incorporated into the liposome beforehand, the amount of the liposome bound to the extracellular matrix can be obtained as radioactivity. Furthermore, to qualitatively judge the binding and covering degree to the extracellular matrix, a labeled antibody for recognizing the integrin  $\alpha 2\beta 1$ -immunoglolublin chimeric protein heterodimer complex on the bound liposome can be combined with a color dye, etc., to dye the portions where the liposome is bound. It is more preferable that the generally used tissue antibody dyeing method is used to use a peroxidase labeled antibody against the integrin  $\alpha 2\beta 1$ -immunoglolublin chimeric protein heterodimer complex and diaminobenzidine in combination, but the measuring method is not limited to it. As a further other method, the area covering the extracellular matrix can be obtained as a covering rate using an image processing analyzer.

Methods for examining the hemostasis of platelets include testing the adhering capability of platelets to the extracellular matrix and the agglutination capability induced by a collagen ("Handbook on the Examination of Blood Coagulation (in Japanese)", p. 65-78, Fukutake, M. and Fujimaki, M. (1987), Uchudo Yagi Shoten, Santro, S.A., Cell, 46, 913-920 (1986), Lethagen, S. and Rugarrn, P., Thrombo Haemost., 67, 185-186 (1982)). Especially the adhering

capability of platelets to the extracellular matrix is an indicator of primary hemostasis. The adhering capability is evaluated by using blood as it is, or platelet rich plasma or platelets washed by a buffer with physiological ions. Therefore, whether or not the integrin  $\alpha 2\beta 1$ -immunoglolublin chimeric protein heterodimer complex liposome obtained in the present invention can be a functional substitute of platelets can be judged in reference its binding capability and the level of the binding capability to the extracellular matrix in the existence of plasma components or at a physiological ion concentration.

If the binding capability of the integrin  $\alpha 2\beta$  1-immunoglolublin chimeric protein heterodimer complex liposome obtained in the present invention to the extracellular matrix in the presence of the plasma components is strong, it suggests that the liposome can be a platelet substitute. Therefore, it can be used as a therapeutic or preventive agent against the congenital and acquired bleeding tendency due to platelet abnormality, and also widely as a platelet transfusion substitute.

Similarly the integrin  $\alpha 2\beta 1$ -immunoglolublin chimeric protein heterodimer complex liposome obtained in the present invention can be a therapeutic or preventive agent for conditions of diseases where vascular endothelial cell disorder is a problem. For example, it was reported that in

the prognosis of PTCA (percutaneous coronary restenosis), the excessive accumulation of platelets on the extracellular matrix exposed by balloon catherter treatment triggers restenosis (Liu, M.W. et al., Circulation, 79, 1374-1378 (1989)). In Example 22, the effect of the integrin  $\alpha 2\beta$ 1-immunoglolublin chimeric protein heterodimer complex liposome to cover the extracellular matrix was confirmed, and this effect can reduce the excessive accumulation of platelets to allow use also as a restenosis preventive. Furthermore, if the integrin  $\alpha 2\beta 1$ -immunoglolublin chimeric protein heterodimer complex liposome is labeled by a medically allowable method, it can be used for monitoring the region of the extracellular matrix exposed by vascular endothelial cell injury, and furthermore, if a drug is enclosed in the liposome, it can also be applied to the targeting therapy for a local injured region.

When any integrin  $\alpha 2\beta 1$ -immunoglolublin chimeric protein heterodimer complex liposome stated in the present invention is used as a platelet substitute, the administration paths include infusion, intravenous administration, etc., and it is usually used by being suspended in any physiologically suitable solution such as a salt solution or plasma, etc. It can be used alone or also in combination with another chimeric protein heterodimer complex with an extracellular matrix receptor or its immunoglobulin. It can also be used together

with another drug containing total platelets. The dose is properly selected to suit the symptom, age, body weight, etc., and can be 0.1 mg to 10 g per day as the amount of the protein for an adult, being able to be administered at a time or in several times. It can also be mixed with a pharmaceutically allowed carrier or excipient, etc., to be applied locally to the injured region as an externally applied drug such as an ointment, liniment or plaster. In this case, the externally applied drug is prepared to be 1 ng/cm² to 1 mg/cm² as the amount of the protein per one time of coating.

## Examples

To describe the present invention in more detail, examples are given below. The general methods of recombinant DNA experiments conformed to those stated in a published book ("Antibody", Harlow, E. and Lane, D. (1988), Cold Spring Harbor Lab. Press, New York).

## Example 1

Construction of human lgG 1 heavy chain expression vector

As human IgG 1 genome gene, a clone identical with reported base sequence information (Ellison, J. W. et al., Nucleic Acids Res., 10, 4071-4079 (1982)) was acquired from a human genomic library (CLONTECH) using a hybridization cDNA probe based on the sequence information. This was used as the template DNA for PCR. As primers for amplifying the DNA fragment containing the hinge region (H) and the constant

region portions (CH2 and CH3) of human IgG1 gene, a DNA oligomer shown in sequence No. 4 of the sequence table (hereinafter a sequence No. of the sequence table is simply called a sequence No.) with BamH I restriction site and a DNA oligomer shown in sequence No. 5 with Xba I restriction site were synthesized.

- 5'-GCGGATCCCGAGCTGCTGGAAGCAGGCTCAG-3' (Sequence No. 4)
- 5' CCTCTAGACGGCCGTCGCACTCATTTA-3' (Sequence No. 5)

The template DNA, primers, dNTPs (an equimolar mixture of dATP, dCTP, dGTP and dTTT) and Taq polymerase (Takara) were mixed in a PCR buffer (100 mM Tris-HCl, 500 mM KCl, 15 mM MgCl<sub>2</sub>, 0.01% gelatin, pH 8.3), and in a thermal cycler (Perkin Elmer Cetus), the mixture was treated at 94 °C for 1 minute for DNA denaturation, at 58°C for 2 minutes for annealing the primers and at 72°C for 3 minutes for elongating the primers. This treatment was performed 30 cycles. The amplified DNA was digested by restriction enzymes BamH I and Xba I, and the DNA fragment was purified by 1% agarose gel according to a general method ("Antibody", Harlow, E. and Lane, D. (1988), Cold Spring Harbor Lab. Press, New York). It was linked, using a T4DNA ligase, with a large DNA fragment of pBluescriptSK(+) (STRATAGENE) purified and digested by restriction enzymes BamH I and Xba I. The plasmid DNA was used to transform Escherichia coli (JM109), and the transformant was selected, to obtain a plasmid DNA (lgG1 Bluescript). Then, expression

vector pcDL-SR  $\alpha$ 296 was digested by restriction enzyme BamH I, and blunted at the termini by T4DNA polymerase treatment, and a Not I linker was linked. The large DNA fragment obtained by digesting it by restriction enzymes Not I and Xho I and the small DNA fragment obtained by digesting IgG1 Bluescript by restriction enzymes Not I and Xho I were purified according to a general method, and linked by T4DNA ligase. It was transformed into Escherichia coli (HB101), and the transformant was selected, to obtain a plasmid DNA. Hereinafter this plasmid (IgG 1SR  $\alpha$ ) is called human IgG1 expression vector. In the following examples, since the basic protocol of gene manipulation is the same as above, the description will be simplified.

## Example 2

Construction of integrin  $\alpha 4$  · IgG heavy chain chimeric protein expression vector

The DNA fragment coding for the extracellular portion of integrin  $\alpha 4$  was obtained by cloning based on reported cDNA sequence information (Takada, Y. et al., EMBO J., 8, 1361-1368 (1989)). The restriction site EcoR I of 1801-base-position of sequence No. 1, the restriction site Stu I of 112-base-position and the restriction site BamH I of 2949-base-position were used for linking the region from the N terminus translation initiation site to Stu I cut site as  $\alpha 4-1$ , the region from Stu I cut site to EcoR I cut site as  $\alpha 4-2$ , and the

region from EcoR I detached site to BamH I detached site as  $\alpha$  4-3. The detailed methods are described below.

The portion coding for  $\alpha 4-1$  was designed to be cloned by linking the DNA oligomers of sequence Nos. 6 to 9, and the DNA oligomers shown in sequence Nos. 6 to 9 were synthesized. For the sequence Nos. 6 and 7, restriction site Xba I was added on the side to code for the N terminus, for linking to a vector. Furthermore, compared with the known sequence information, the bases at the 60-, 63- and 64-positions were substituted from C to T, C to A and C to G respectively, and the bases at the 112- and 114-positions were substituted freeom C to A and C to G respectively. Because of substitution at the 112- and 114position, restriction site Stu I was inserted on the side to code for the N terminus of sequence Nos. 8 and 9. The 5' termini of the synthesized oligomers were phosphated and annealed, and were linked using T4DNA ligase. After completion of linking, restriction enzymes Xba I and Stu I were used for cutting, and electrophoresis was effected by 5% agarose (NuSieve GTGagarose, FMC) gel. The intended DNA fragment  $(\alpha 4-1)$  of about 120 bp was cut out and purified.

<sup>5&#</sup>x27; - CTAGACCACCATGTTCCCCACCGAGAGCGCATGGCTTGGGAAGCGAGGCGCGAACCCGGGCCCCGGA
GCTGCA-3'
(Sequence No. 6)

<sup>5&#</sup>x27;--GCTTCGGGGCCCGGGTTCGCGCCTCGCTTCCCAAGCCATGCGCTCTCGGTGGGGAACATGGTGGT-3'
(Sequence No. 7)

<sup>5&#</sup>x27; - CTCCGGGAGACGGTGATGCTGTTGCTGTGCCTGGGGGTCCCGACCGGCAGG-3'

5' - CCTGCCGGTCGGGACCCCCAGGCACAGCATCACCGTCTCCCGGAGTCGA-3'
(Sequence No. 9)

Then, the RNA of human osteosarcoma cell line MG63 (ATCC CRL 1427) as an integrin  $\alpha 4$  expressing cell was separated, and PolyA(+)RNA was purified using oligo dT cellulose column (NEB). Based on it, a single stranded cDNA was synthesized using a reverse transcriptase (GIBCO), and used as the template for PCR. As primers for amplifying  $\alpha 4-2$  and  $\alpha 4-3$  DNAs, four DNA oligomers of sequence Nos. 10 to 13 with Pst I and Stu I restriction sites inserted (sequence No. 10) or BamH I restriction site inserted (sequence No. 13) were synthesized.

5' - CACTGCAGGCAGGCCTTACAACGTGGACACTGAGAGC-3' (Sequence No. 10)

5' - GCAGAAACCTGTAAATCAGCAG-3' (Sequence No. 11)

5'- GCATTTATGCGGAAAGATGTGC-3' (Sequence No. 12)

5' - CGGGATCCGTGAAATAACGTTTGGGTCTT-3' (Sequence No. 13)

The template cDNA, primers, dNTPs and Taq polymerase were mixed in a PCR buffer, and the mixture was treated by a thermal cycler at 94°C for 1 minute for DNA denaturation, at 58°C for 2 minutes for annealing the primers and at 72°C for 3 minutes for elongating the primers. This treatment was performed 30 cycles. The amplified DNA fragments of  $\alpha$ 4-2 and  $\alpha$ 4-3 were digested by Pst 1 and EcoR 1 respectively, or EcoR 1

and BamH I and sub-cloned into pBluescriptKS(+) (STRATAGENE), to prepare plasmid DNAs (hereinafter called  $\alpha 4-2$  Bluescript and  $\alpha 4-3$  Bluescript). Then, upstream of the  $\alpha 4-2$  Bluescript,  $\alpha 4-1$  was linked using Xba I and Stu I restriction sites, to prepare a plasmid DNA (hereinafter called  $\alpha 4-1-2$  Bluescript).

The  $\alpha 4$ -1-2 Bluescript was digested by restriction enzyme Not I, and blunted at the termini by T4DNA polymerase treatment, being digested by restriction enzyme EcoR I, to prepare a small DNA fragment. The  $\alpha 4$ -3 Bluescript was digested by restriction enzymes EcoR I and BamH I, to prepare a small DNA fragment. The two small DNA fragments were simultaneously linked to a large DNA fragment obtained by digesting  $lgG_1SR\alpha$  by restriction enzymes EcoR V and BamH I, to obtain a plasmid DNA. The obtained base sequence coding for integrin  $\alpha 4 \cdot lgG$  heavy chain chimeric protein is shown as sequence No. 1. The plasmid (integrin  $\alpha 4 \cdot lgGSR\alpha$ ) is hereinafter called integrin  $\alpha 4 \cdot lgG$  heavy chain chimeric protein expression vector.

Example 3

Construction of  $\beta$ 1 - IgG heavy chain chimeric protein expression vector

The RNA of human fibroblast cell line MRC5 (ATCC CCL 171) as an integrin  $\beta$ 1 expressing cell was separated, and oligo dT cellulose column was used to purify PolyA (+)RNA. Based on it, a single stranded cDNA was synthesized using a reverse

transcriptase, and used as the template for PCR. As primers, two DNA oligomers of sequence Nos. 14 and 15 with BamH I site (sequence No. 15) inserted on the side coding for C terminus were synthesized according to the sequence information (Cott Argraves, W. et al., J. Cell Biol., 105, 1183-1190 (1987)).

5'-GCGGAAAAGATGAATTTACAAC-3' (Sequence No. 14)

5'-GTGGGATCCTCTGGACCAGTGGGACAC-3' (Sequence No. 15)

The template cDNA, primers, dNTPs and Taq polymerase were mixed in a PCR buffer, and treated by a thermal cycler at 94°C for 1 minute for DNA denaturation, at  $57^{\circ}$ C for 2 minutes for annealing the primers and at 72°C for 3 minutes for elongating the primers. This treatment was performed 30 cycles. The amplified DNA was blunted at the termini by T4DNA polymerase treatment, and digested by restriction enzyme BamH 1. the DNA fragment was purified. Subsequently, the DNA fragment obtained in PCR before was sub-cloned at the Sma I and BamH ! sites of pBluescriptKS(+). A small DNA fragment purified by digesting it by restriction enzymes EcoR I and BamH I was inserted into a large DNA fragment of  $IgG_1SR\alpha$  treated by restriction enzymes EcoR I and BamH I, to obtain a plasmid The obtained base sequence coding for  $\beta$ 1 · lgG heavy DNA. chain chimeric protein is shown in sequence No. 2. plasmid (integrin  $\beta$ 1 · lgGSR $\alpha$ ) is hereinafter called integrin B1 · IgG heavy chain chimeric protein expression vector. Example 4

Transfection of  $\alpha 4 \cdot \lg G$  heavy chain chimeric protein expression vector and  $\beta 1 \cdot \lg G$  heavy chain chimeric protein expression vector into animal cells, and their expression

Integrin  $\beta$ 1 · IgGSR $\alpha$  as  $\beta$ 1 · IgG heavy chain chimeric protein expression vector and pSV2dhfr (BRL) were mixed at a ratio of 10 : 1, and the mixture and lipofectin reagent (GIBCO BRL) were gently mixed and allowed to stand at room temperature for 15 minutes. The mixture was added dropwise to dihydrofolic acid reductase deficient CHO cells (ATCC CRL 9096). After 18 hours of dropwise addition, the mixture was cultured in a medium (10%FBS (GIBCO), nucleic acid-containing  $\alpha$ MEM medium (GIBCO BRL)) for about 2 days, and the cells were dispersed by trypsin-EDTA treatment. The cells were suspended in a first selective medium (10% FBS-containing nucleic acidfree  $\alpha$ MEM medium (GIBCO BRL)), and the suspension was disseminated into a 96-well plate (CORNING), for selective culture for about 10 days. Then, the amount of integrin  $\beta$ 1. IgG heavy chain chimeric protein produced in the culture supernatant was determined according to the ELISA method (described later), and the clone showing the highest production was stabilized by cloning according to the limiting dilution method.

Then, into the stabilized integrin  $\beta$ 1 - IgG heavy chain chimeric protein producing CHO cells, the integrin  $\alpha$ 4 - IgG heavy chain chimeric protein expression vector was transfected

according to the lipofectin method as described before. is, integrin  $\alpha 4 \cdot lgGSR\alpha$  and pSV2neo (BRL) were mixed at 10 : 1, and the mixture was mixed with lipofectin reagent. The mixture was added dropwise into the cells. After 18 hours of dropwise addition, the mixture was cultured in the said first selective medium for about 2 hours, and the cells were dispersed by trypsin-EDTA treatment. The cells were suspended in a second selective medium (nucleic acid-free lphaMEM medium (GIBCO BRL) containing 10% FBS (GIBCO) and 1 mg/ml neomycin (GIBCO)), and on a 96-well plate (CORNING), resistant cells were selectively cultured for about 10 days. The amount of integrin  $\alpha 4 \cdot lgG$  heavy chain chimeric protein and the amount of integrin \$1 - IgG heavy chain chimeric protein produced in the culture supernatant were determined according to the ELISA method (described later), and a clone which produced both the chimeric proteins by almost the same amounts was picked up. The clone was cloned twice according to the limiting dilution method, to be stabilized as a clone capable of producing lpha 4 . IgG heavy chain-Bl·lgG heavy chain chimeric protein heterodimer complex.

Example 5

Determination of produced integrin  $\alpha 4$  - IgG heavy chain chimeric protein and integrin  $\beta 1$  - IgG heavy chain chimeric protein by the ELISA method

Fifty microliter per well of anti-human integrin  $\alpha 4$ 

antibody (Becton & Dickinson, Clone L25.3) or anti-human integrin  $\beta$ 1 antibody (Coulter, Clone 4B4)(12  $\mu$ g/ml each) was put into a 96-well immunoplate (NUNC), and allowed to stand at 4°C for 16 hours. Then, each well was washed by Dulbecco's phosphate buffered saline (Nissui Seiyaku, not containing Ca or Mg ions, hereinafter called PBS(-)) twice, and non-specific reaction was blocked by PBS(-) containing 25% Block Ace (Snow Brand Milk Products Co., Ltd.). After blocking, the culture supernatant of CHO cells grown in selective medium was properly diluted, and reacted with the coated antibody at room temperature for 1 hour. After the reaction, the surface of the plate was washed with 0.02% Tween-containing PBS(-) (hereinafter called T-PBS) twice. It was then caused to react with biotinated anti-human IgG antibody (Vector) for 1 hour, and the reaction mixture was washed with T-PBS twice, and in succession caused to react with avidin-horseradish peroxidase (Sigma) for 1 hour. The reaction mixture was washed with PBS(-) twice. The PBS(-) was perfectly aspirated, and orthophenylenediamine was used as a substrate for color development. The absorbance at 490 nm were measured using a microplate reader (Bio-rad NOVAPATH), and the clone showing a high absorbance value was selected.

Example 6

Purification of  $\alpha 4 \cdot \text{lgG}$  heavy chain- $\beta 1 \cdot \text{lgG}$  heavy chain chimeric protein heterodimer complex

(1) Culture of CHO cells and preparation of cultured supernatant solution

(2) Protein A column chromatography

The starting solution for further purification was passed through Prosep Guard column (bioPROCESSING), and applied to Prosep A column (bioPROCESSING). After completion of application, it was washed with 10 times the column volume of PBS(-), and the proteins were eluted at a pH 6 3 gradient of 0.1M citrate buffer solutions. The peak fraction eluted at pH 3 was collected, and 1M Tris-HCl solution (pH 8.5) was added by 0.1 volume for neutralization. The solution was dialyzed against PBS(-).

(3) Affinity column chromatography

FMP activated Cellulofine (Seikagaku Kogyo) was equilibrated by a coupling buffer (50 mM Na<sub>2</sub>CO<sub>3</sub>-NaHCO<sub>3</sub> pH 8.5), and a peptide showing sequence No. 3 (hereinafter called CS-1 peptide) synthesized by a peptide synthesizer was added. mixture was inverted and mixed at 4 °C for 16 hours. Cys Leu His Gly Pro Glu Ile Leu Asp Val Pro Ser Thr (Sequence No. 3) After completion of mixing, the mixture was washed with the coupling buffer, and a blocking buffer (0.1 mM monoethanolamine, 50 mM Tris-HCI, pH 8.0) was added. mixture was inverted and mixed further at room temperature for 6 hours. Then, the mixture was suffificiently washed with TBS solution (150 mM NaCl, 20 mM Tris-HCl, 1 mM MnCl<sub>2</sub>, pH 7.5), to prepare CS-1 peptide bound Cellulofine column. To the column the starting solution for further purification was applied and allowed to stand at room temperature for 3 hours, and washed with 10 times the column volume of a washing buffer (1M NaCl, 0.1% Triton, 20 mM Tris-HCl, 1 mM MnCl 2, pH 7.5) and the same volume of the TBS solution. After completion of washing, an elution buffer (10 mM EDTA, 150 mM NaCl, 20 mM Tris-HCl, pH 7.5) was used, to elute the proteins bound to the CS-1 column. The eluate was collected and dialyzed against PBS(-).

## (4) SDS-PAGE

The eluted fractions of (3) were subjected to SDS-PAGE under non-reducing or reducing condition using 6.0 or 7.0% acrylamide gel, and the gel was stained with Coomassie-blue.

As a result, under non-reducing condition, two bands considered to be attributable to the  $\alpha 4$  -lgG heavy chain- $\beta 1$  -IgG heavy chain chimeric protein heterodimer complex and its polymer were observed. Under reducing condition, two bands (170 kDa and 135 kDa) considered to be attributable to the integrin  $lpha extsf{4} \cdot extsf{IgG}$  heavy chain chimeric protein and the integrin eta1 - IgG heavy chain chimeric protein and two bands (80 kDa and 90 kDa) considered to be attributable to the intramolecular cleavage of the integrin  $\alpha 4 \cdot \lg G$  heavy chain chimeric protein (Hemler, M. E. et al., J. Biol. Chem., 262, 11478-11485 (1987)) were observed. These results suggest that the eluted protein of (3) has a molecular structure considered to be lpha 4 . lgG heavy chain-β1·lgG heavy chain chimeric protein heterodimer complex, and that the molecules constituting the heterodimer are linked by a disulfide bond between the IgG heavy chains.

Example 7

Identification of  $\alpha 4 \cdot \text{IgG}$  heavy chain—  $\beta 1 \cdot \text{IgG}$  heavy chain chimeric protein heterodimer complex, and examination of its structural stability

(1) Immunoprecipitation using anti-integrin antibodies and influence of a cationic chelating agent

The basic method conformed to a published book

("Antibodies", Harlow, E. et al., (1988), Cold Spring Harbor

Lab. Press, New York). That is, the eluted pfotein of Example

6 (3) considered to be  $\alpha$ 4 - lgG heavy chain- $\beta$ 1 - lgG heavy chain chimeric protein heterodimer complex was \$^{125}I-labeled using the lactoperoxidase method. Then, Affigel-10 (Bio-rad) was washed with 0.1 M Hepes solution (pH 8.0), and normal murine IgG, anti-human integrin  $\alpha 4$  antibody (clone 11C2B) and anti-human integrin  $\beta$ 1 antibody (clone 4B4) were added. Reaction was effected at 4°C for 16 hours to cause covalent bonding, to prepare normal murine lgG beads and the respective antibody Then, the  $^{125}$ l labeled  $\alpha 4 \cdot \lg G$  heavy chain- $\beta 1 \cdot \lg G$ beads. heavy chain chimeric protein heterodimer complex and normal murine IgG beads were inverted and mixed at 4 °C for 4 hours for preclearing, and the mixture and the antibody beads were inverted and mixed at 4°C for 16 hours. After completion of mixing, the beads were washed with a washing buffer (200 mM Tris-HCI, 0.5 M NaCI, 0.1% NP-40, 1 mM MgCl<sub>2</sub> or 10 mM EDTA, pH 8.0) three times. After completion of washing, a sample buffer for electrophoresis was added to the beads for treatment at 100 ℃ for 5 minutes, and the mixture was centrifuged. The supernatant solution was analysed by electrophoresis under reducing condition. After completion of electrophoresis, the gel was dried by a gel dryer, and the protein was detected by autoradiography.

As a result of immunoprecipitation in the presence of 1 mM MgCl2, from the beads of both the anti-human integrin  $\alpha 4$  antibody and the anti-human integrin  $\beta 1$  antibody, the same

precipitation patterns expected from the structure of  $\alpha 4 \cdot \lg G$  heavy chain- $\beta 1 \cdot \lg G$  heavy chain chimeric protein heterodimer complex were obtained. Thus, the protein obtained in (3) of Example 6 was identified as  $\alpha 4 \cdot \lg G$  heavy chain- $\beta 1 \cdot \lg G$  heavy chain chimeric protein heterodimer complex.

On the other hand, the immunoprecipitation pattern obtained by using the anti-integrin  $\beta$ 1 antibody beads in the presence of 10 mM EDTA was the same as that in the presence of 1 mM MgCl<sub>2</sub>, to clarify that the association between integrin  $\alpha$ 4·lgG heavy chain chimeric protein and integrin  $\beta$ 1·lgG heavy chain chimeric protein does not depend on cations. The above results suggest that the eluted protein obtained in (3) of Example 6 was certain  $\alpha$ 4·lgG heavy chain- $\beta$ 1·lgG heavy chain chimeric protein heterodimer complex, and if the result of (4) of Example 6 is also taken into account, it is strongly suggested that the association between both the proteins is stable association through a disulfide bond existing the lgG heavy chains.

(2) Examination on the structural stability of  $\alpha 4 \cdot \lg G$  heavy chain  $-\beta 1 \cdot \lg G$  heavy chain chimeric protein heterodimer complex by sequential immunoprecipitation

According to (1),  $^{125}$ l labeled  $\alpha 4 \cdot \text{lgG}$  heavy chain- $\beta 1 \cdot \text{lgG}$  heavy chain chimeric protein heterodimer complex was caused to react with normal murine lgG beads, anti-human integrin  $\alpha 4$  antibody (11C2B) beads or anti-human integrin  $\beta 1$  antibody

(484) beads at  $4^{\circ}$ C for 4 hours, and the reaction mixture was washed. After washing, the reaction mixture was boiled at  $100^{\circ}$ C for 5 minutes in the presence of 2% SDS and centrifuged, and the supernatant (primary immunoprecipitation sample) was diluted to 10 times by 1% BSA-containing PBS, and was again reacted with the anti-integrin  $\beta$ 1 antibody beads and the anti-integrin  $\alpha$ 4 antibody beads at  $4^{\circ}$ C for 16 hours. After completion of reaction, the beads were washed, and a sample buffer for electrophoresis was added. The mixture was treated at  $100^{\circ}$ C for 5 minutes and centrifuged. The supernatant solution (secondary immunoprecipitation sample) was analyzed by SDS-PAGE/autoradiography.

As a result, the electrophoretic pattern obtained by the primary immunoprecipitation was also similarly observed in the secondary immunoprecipitation. This result suggests that the association between the  $\alpha 4 \cdot \lg G$  heavy chain chimeric protein and the  $\beta 1 \cdot \lg G$  heavy chain chimeric protein in the  $\alpha 4 \cdot \lg G$  heavy chain- $\beta 1 \cdot \lg G$  heavy chain chimeric protein heterodimer complex is not dissociated either by boiling in the presence of 2% SDS, and strongly supports that the complex has a stable heterodimer structure based on a disulfide bond.

Example 8

Binding of  $\alpha 4 \cdot \text{IgG}$  heavy chain  $-\beta 1 \cdot \text{IgG}$  heavy chain chimeric protein heterodimer complex to VCAM-1

It was examined that the  $\alpha$ 4-lgG heavy chain- $\beta$ 1-lgG

heavy chain chimeric protein heterodimer complex produced by CHO cells can be bound to the ligand of integrin  $\alpha 4 \, \beta \, 1$  by using the cells expressing VCAM-1. Human normal umbilical intravenous endothelial cells were cultured with IL-1 3U/ml for 16 hours, to prepare VCAM-1 expressing cells. The cells dispersion as single cells. The cells (2 x  $10^5$  cells per sample tube) were cultured with the supernatant of the CHO cells producing  $\alpha 4 \cdot \lg G$  heavy chain- $\beta 1 \cdot \lg G$  heavy chain chimeric protein heterodimer complex for 30 minutes in the presence of 1 mM (final concentration) MnCl<sub>2</sub> or 3 mM (final concentration) EDTA. After completion of reaction, the cells were washed twice by centrifugation at 1200 rpm at room temperature for 5 minutes using a buffer for binding assay(24 mM Tris-CHI, 10 mM Hepes, 150 mM NaCI, 1 mM MnCI 2 or 1 mM EDTA, 1% BSA, 2 mM glucose, pH 7.4). After washing, FITC labeledanti-human IgG antibody (Cappel) was added, and incubated at room temperature for 20 minutes. The cells were washed by the same buffer, and the chimeric proteins bound to the cells were determined by a flow cytometer (ELITE, Coulter).

The results are shown in Fig. 1. It was observed that the fluorescence intensity showing the binding of  $\alpha 4 \cdot \lg G$  heavy chain- $\beta 1 \cdot \lg G$  heavy chain chimeric protein heterodimer complex increased by culturing the VCAM-1 expression cells with the supernatant containing  $\alpha 4 \cdot \lg G$  heavy chain- $\beta 1 \cdot \lg G$  heavy chain

chimeric protein heterodimer complex. The binding was inhibited by adding anti-human integrin antibodies (anti- $\alpha$ -antibody: clone L25.3, 10  $\mu$ m/ml + anti- $\beta$ 1 antibody: clone 4B4, 10  $\mu$ m/ml) or 3 mM EDTA. This result suggests that the  $\alpha$ 4·lgG heavy chain- $\beta$ 1·lgG heavy chain chimeric protein heterodimer complex can be bound to VCAM-1 like the integrin  $\alpha$ 4 $\beta$ 1 existing on the surfaces of cell membranes, and furthermore that the binding is  $\alpha$ 4 $\beta$ 1-specific and retains a feature of the binding that it is dependent on cations. Example 9

Binding of  $\alpha 4 \cdot \text{lgG}$  heavy chain- $\beta 1 \cdot \text{lgG}$  heavy chain chimeric protein heterodimer complex to a peptide fragment of fibronectin

The capability of  $\alpha 4 \cdot \lg G$  heavy chain- $\beta 1 \cdot \lg G$  heavy chain chimeric protein heterodimer complex to be bound to the peptide fragment (sequence No. 3) of the other ligand, fibronectin was also examined.

At first, according to the said report (Humphries, M. J. et al., J. Biol. Chem., 262, 6886-6892 (1987)), the peptide fragment of sequence No. 3 (CS-1 peptide) was bound to rabbit lgG (Sigma), to prepare CS-1-lgG. The CS-1-lgG was diluted by PBS(-), and put in a 96-well immunoplate (NUNC) by 100  $\mu$  l/well, and allowed to stand at 4°C for 16 hours, to be formed as a solid phase on the plate.

After completion of standing, the surface of the plate was

washed with PBS(-) twice and treated with denatured 1% BSA(heat-natured at 80℃ for 10 minutes)-PBS solution ( 300  $\mu$ I/well) at 4°C for 3 hours to block the nonspecific reaction. Then, the solid phase CS-1-lgG and the CHO culture supernatant (100  $\mu$ I) containing  $\alpha$ 4  $\cdot$  lgG heavy chain- $\beta$ 1  $\cdot$  lgG heavy chain chimeric protein heterodimer complex were reacted with each other at 30°C for 3 hours. The non-bound α4·IgG heavy chain- $\beta$ 1 - IgG heavy chain chimeric protein heterodimer complex was removed by washing with 0.1% BSA-containing TBS buffer (150 mM NaCl, 25 mM Tris-HCl, 1 mM MnCl<sub>2</sub>, pH 7.4) twice, and the bound  $\alpha$ 4 • lgG heavy chain- $\beta$ 1 • lgG heavy chain chimeric protein heterodimer complex was detected by biotin labeled anti-human IgG antibody (Vector) as the primary antibody and avidin labeled horseradish peroxidase (Sigma) as the secondary antibody. The surface of the plate was washed with the TBS Orthophenylenediamine was added as a substrate to it for color development, and the absorbance at 490 nm were measured.

The results are shown in Fig. 2. The reaction with  $\alpha 4 \cdot 1$  IgG heavy chain- $\beta 1 \cdot 1$ gG heavy chain chimeric protein heterodimer complex showed a rise in the absorbance indicating the binding to CS-1 peptide. The binding was almost perfectly inhibited by the presence of anti-integrin  $\alpha A$  antibody (clone L25.3), anti-integrin  $\beta 1$  antibody (clone 4B4) or 5 mM EDTA. Therefore, it was clarified that  $\alpha 4 \cdot 1$ gG heavy chain- $\beta 1 \cdot 1$ gG

heavy chain chimeric protein heterodimer complex can be bound also to the CS-1 peptide which is a peptide fragment of fibronectin, and that a feature of binding that it depends on cations is retained.

## Example 10

Evaluation of an inhibitory peptide by using a system for determining the binding of  $\alpha 4 \cdot \lg G$  heavy chain- $\beta 1 \cdot \lg G$  heavy chain chimeric protein heterodimer complex to a peptide fragment of fibronectin

In the binding determination system of Example 9, the effects of three peptides, i.e., sequence No. 16 (hereinafter called GPEILDVPST), 17 (hereinafter called GPEILEVPST) and 18 (hereinafter called GRGDSP) were examined.

Gly Pro Glu Ile Leu Asp Val Pro Ser Thr

Gly Pro Glu Ile Leu Glu Val Pro Ser Thr

Gly Arg Gly Asp Ser Pro

(Sequence No. 16)

(Sequence No. 17)

The all peptides were synthesized by a peptide synthesizer. The peptide and 100  $\mu$ l of CHO cultured supernatant solution containing  $\alpha\alpha 4 \cdot \lg G$  heavy chain- $\beta 1 \cdot \lg G$  heavy chain chimeric protein heterodimer complex were mixed at room temperature for 20 minutes, and the binding to CS-1- $\lg G$  was determined according to the method in Example 9. The results are shown in Fig. 3. GPEILDVPST showed temperaturedependent inhibitory activity in a range of 0.1 to 10  $\mu$ mg/ml, but GPEILEVPST and GRGDSP did not show any inhibition of the

binding. These results show that the binding determination system in Example 9 allows to detect the inhibiting effect of the peptide (GPEILDVPST) inhibiting the binding between integrin  $\alpha 4 \beta 1$  and CS-1 peptide specifically .

#### Example 11

Construction of integrin  $\alpha 2$  –  $\lg G$  heavy chain chimeric protein expression vector

The DNA fragment coding for the extracellular portion of integrin  $\alpha 2$  was divided into  $\alpha 2-1$  and  $\alpha 2-2$  based on the reported cDNA sequence information (Takada, Y. et al., J. Cell. Biol., 109, 397-407 (1989)) and subcloned, and they were integrated on an expression vector. At first, the RNA of human fibroblast cell line MRC-5 (ATCC CCL 171) as integrin  $\alpha 2$  expressing cell was separated, and an oligo dT cellulose column was used to purify PolyA(+)RNA. Based on it, a single stranged cDNA was synthesized and used as the template of PCR. As PCR primers, DNA oligomers of sequence Nos. 20 and 21 were synthesized for  $\alpha 2-1$ , and DNA oligomers of sequence Nos. 22 and 23, for  $\alpha 2-2$ .

5'	-GCTCGAGCAAACCCAGCGCAACTACGG-3'	(Sequence	No.	20)
5'	-ATAGTGCCCTGATGACCATTG-3'	(Sequence	No.	21)
5'	-GATGGCTTTAATGATGTGATTG-3'	(Sequence	No.	22)
5'	-TGTTGGTACTTCGGCTTTCTC-3'	(Sequence	No.	23)

The template cDNA, primers, dNTPs and Taq polymerase were mixed in a PCR buffer and PCR was performed 30 cycles by a

thermal cycler (reaction conditions: 94℃ 1 minute - 60℃ 2 minutes -  $72^{\circ}$ C 3 minutes). The amplified DNA fragment of  $\alpha 2-1$ was digsted by restriction enzymes Xho I and EcoR I , and the DNA fragment of  $\alpha 2-2$  was blunted at the termini by T4DNA polymerase treatment and digested by restriction enzyme EcoR 1. Each fragment was purified. The two purified DNA fragments were caused to react in a phosphating reaction solution (50 mM Tris-HCl, 10 mM MgCl<sub>2</sub>, 25 mM DTT, 1 mM ATP, 0.1 U/ $\mu$ l T4 polynucleotide kinase (Takara), pH 8.0) at 37 °C for 1 hour, and the reaction mixture was heat-treated at 68℃ for 5 minutes to inactivate the enzyme. Then,  $lgG_1SR\alpha$  prepared in Example 1 was digested by restriction enzyme BamH I and caused to react in Klenow reaction solution (66 mM Tris-HCl, 10 mM MgCl 2, 10 mM DTT, 0.2 mM dNTPs, 0.05 U/  $\mu$ I Klenow fragment (Takara), pH 7.5) at 37°C for 30 minutes, to blunt the termini, and the reaction mixture was heat-treated at 70°C for 5 minutes to inactivate Furthermore, a large DNA fragment was digested by the enzyme. restriction enzyme Xho I, and pudified. The two ( $\alpha 2-1$  and  $\alpha$ 2-2) DNA fragments phosphated before were inserted into the large DNA fragment, to obtain a plasmid DNA. The obtained base sequence coding for integrin  $\alpha 2 \cdot \lg G$  heavy chain chimeric protein is shown in sequence No. 19. This plasmid (integrin lpha $2 \cdot \lg GSR \alpha$ ) is hereinafter called  $\alpha 2 \cdot \lg G$  heavy chain chimeric protein expression vector.

Example 12

Transfection of integrin  $\alpha 2 \cdot \text{IgG}$  heavy chain chimeric protein expression vector and integrin  $\beta 1 \cdot \text{IgG}$  heavy chain chimeric protein expression vector into animal cells, and their manifestation

The integrin  $\alpha 2 \cdot \lg G$  heavy chain chimeric protein expression vector was transfected into the integrin eta1  $\cdot$ 1  $\circ$ 1  $\circ$ 1 heavy chain chimeric protein producing CHO cells prepared and stabilized in Example 4, according to the lipofectin method described in Example 4. That is, integrin  $\alpha 2 \cdot IgGSR\alpha$  and pSV2neo (BRL) were mixed at 10 : 1, and the mixture was mixed with lipofectin reagent. The mixture was added dropwise to the cells. Eighteen hours after completion of dropwise addition, the mixture was cultured in a first selective medium for 2 days, and the cells were dispersed by trypsin-EDTA treatment. The cells were suspended in a second selective medium, and the suspension was disseminated into a 96-well plate. Resistant cells were selectively cultured for about 10 days. Then, the amount of integrin  $\alpha$ 2  $\cdot$  lgG heavy chain chimeric protein and the amount of integrin \$1 - lgG heavy chain chimeric protein produced in the culture supernatant were determined according to the ELISA method (described later), and a clone producing almost the same amounts of both the chimeric proteins was picked up. The clone was cloned twice according to the limiting dilution analysis, to be stabilized as a clone capable of producing lpha

 $2 \cdot \lg G$  heavy chain- $\beta 1 \cdot \lg G$  heavy chain chimeric protein heterodimer complex.

Example 13

Determination of the amounts of integrin  $\alpha 2 \cdot \lg G$  heavy chain chimeric protein and integrin  $\beta 1 \cdot \lg G$  heavy chain chimeric protein by the ELISA method

Fifty microliter per well of anti-human integrin lpha 2antibody (Becton & Dickinson, clone P1E6) or anti-human integrin  $\beta$ 1 antibody (clone 4B4)( $2\mu g/ml$  each) was put into a 96-well immunoplate, and allowed to stand at  $4^{\circ}$ C for 16 hours. Then, each well was washed with PBS(-) twice, blocked, and the culture supernatant of the CHO cells grown in second selective medium was properly diluted and reacted with the coated-antibody at room temperature for 1 hour. After the reaction, the surface of the plate was washed with T-PBS twice, and caused to react with biotinated anti-human IgG antibody for 1 hour and with avidin-horseradish peroxidase for 1 hour, and the reaction mixture was washed with PBS(-) twice. After completion of reaction, orthophenylenediamine was used as a substrate for color development, and the absorbance values at 490 nm were measured using a microplate reader. clone showing a high absorbance value was selected.

Example 14

Purification of  $\alpha 2 \cdot \text{lgG}$  heavy chain- $\beta 1 \cdot \text{lgG}$  heavy chain chimeric protein heterodimer complex

(1) Culture of CHO cells and preparation of cultured supernatant solution

The CHO cells highly capable of producing  $\alpha 2 \cdot \text{lgG}$  heavy chain- $\beta 1 \cdot \text{lgG}$  heavy chain chimeric protein heterodimer complex were cultured in an  $\alpha \text{MEM}(-)$  medium containing 5% FBS (Ultralow lgG grade) for 1 day, to reach semiconfluent, and they were cultured on an  $\alpha \text{MEM}(-)$  medium containing 1%FBS (Ultra-low lgG grade) for 3 days. The culture supernatant was collected, and concentrated to 1/10 volume by ultrafiltration. Then, 1M Hepes solution (pH 8.0) was added to achieve a final concentration of 5 mM, to obtain a starting solution for further purification.

### (2) Protein A colum chromatography

The starting solution for further purification was passed through Prosep Guard column, and applied to Prosep A column. After completion of application, it was washed with 10 times the column volume of PBS (-), and in succession, the proteins were eluted at a pH 6 to 3 gradient of 0.1M citrate buffers. The peak fraction eluted at pH 3 was collected, and 1M Tris-HCl solution (pH 8.5) was added by 0.1 volume for neutralization. The mixture was dialyzed against PBS(-).

#### (3) Affinity column chromatography

According to a report (Kirchhofer, D. et al., J. Biol. Chem., 265, 615-618 (1990)), a collagen immobilized column with a collagen (Typel, Sigma) coupled to cyanogen-bromide-

activated Sepharose (Sigma) was prepared. Then, the starting solution for further purification was equilibrated in a TBS buffer (150 mM NaCl, 50 mM Tris-HCl, 1 mM MgCl<sub>2</sub>, 1 mM MnCl<sub>2</sub>, pH 7.5), applied to a column, allowed to stand at room temperature for 3 hours, and washed with 10 times the column volume of a washing buffer (150 mM NaCl, 50 mM Tris-HCl, 1 mM MgCl<sub>2</sub>, 1 mM MnCl<sub>2</sub>, 100 mM octyl glucopyranoside, pH 7.5). After completion of washing, an elution buffer (20 mM EDTA, 150 mM NaCl, 50 mM Tris-HCl, 50 mM octyl glucopyranoside, pH 7.5) was used to elute the protein bound to the column. The eluate was collected and dialyzed against PBS(-).

## (4) SDS-PAGE

The eluted fraction of (3) was subjected to SDS-PAGE using 7.0% acrylamide gel under non-reducing or under reudcing condition, and the gel was stained with Coummassie-blue. As a result, a band considered to be attributable to  $\alpha 2 \cdot \lg G$  heavy chain- $\beta 1 \cdot \lg G$  heavy chain chimeric protein heterodimer complex was observed. Under reducing condition, two bands (185 kDa and 135 kDa) considered to be attributable to integrin  $\alpha 2 \cdot \lg G$  heavy chain chimeric protein and integrin  $\beta 1 \cdot \lg G$  heavy chain chimeric protein were observed. These results suggest that the eluted protein has a molecular structure considered to be  $\alpha \alpha 2 \cdot \lg G$  heavy chain- $\beta 1 \cdot \lg G$  heavy chain chimeric protein heterodimer complex, and is linked by a disulfide bond between the  $\log G$  heavy chains.

# Example 15

Identification of  $\alpha 2$  - IgG heavy chain-  $\beta 1$  - IgG heavy chain chimeric protein heterodimer complex, and examination of its structural stability

The eluted protein of (3) of Example 14 was  $^{125}$ l-labeled, and subjected to immunoprecipitation using the beads coupled with normal murine lgG, anti-human integrin  $\alpha$ 2 antibody (clone P1E6) or anti-human integrin  $\beta$ 1 antibody (clone 4B4) as described in Example 7, and to SDS-PAGE/autoradiography under reducing condition.

As a result, in both 1 mM MgCl<sub>2</sub> and 10 mM EDTA, from the beads of both anti-human integrin  $\alpha$ 2 antibody and anti-human integrin  $\beta$ 1 antibody, the same precipitation patterns expected from the structure of  $\alpha$ 2·lgG heavy chain- $\beta$ 1·lgG heavy chain chimeric protein heterodimer complex could be obtained. These results show that the eluted protein obtained in (3) of Example 14 is certainly  $\alpha$ 2·lgG heavy chain- $\beta$ 1·lgG heavy chain chimeric protein heterodimer complex, and with the results of (4) of Example 14 also taken into account, it is strongly suggested that the association of both the proteins is stable through a disulfide bond existing the lgG heavy chains.

#### Example 16

Examination on the capability of  $\alpha 2 \cdot \text{lgG}$  heavy chain- $\beta 1 \cdot \text{lgG}$  heavy chain chimeric protein heterodimer complex to be bound

to a collagen, and its specificity

The capability of  $\alpha 2 \cdot \lg G$  heavy chain- $\beta 1 \cdot \lg G$  heavy chain chimeric protein heterodimer complex to be bound to a collagen which is a ligand of integrin  $\alpha 2\beta 1$  was examined.

At first, a collagen (Cell Matrix Typel 3 mg/ml) was diluted to 0.1  $\mu$ g/ml by 0.02M acetic acid solution, and put in an immunoplate by 100  $\mu$ l/well, being kept at 4°C for 16 hours. Then, the collagen solution was removed by suction, and the plate was washed with PBS(-) twice for neutralization. Heat-denaturated 1% BSA-PBS solution was put in the plate by 300  $\mu$  l/well for blocking at room temperature for 3 hours. After completion of blocking, it was rinsed with PBS(-) twice, to prepare a collagen coated plate.

The cultured supernatant of CHO (100  $\mu$ I) containing  $\alpha$ 2·lgG heavy chain- $\beta$ 1·lgG heavy chain chimeric protein heterodimer complex was reacted at 30°C for 3 hours. After completion of reaction, as described in Example 9, the amount of bound  $\alpha$ 2·lgG heavy chain- $\beta$ 1·lgG heavy chain chimeric protein heterodimer complex was determined.

As a result, as shown in Fig. 4, the absorbance showing the binding of  $\alpha 2 \cdot \lg G$  heavy chain- $\beta 1 \cdot \lg G$  heavy chain chimeric protein heterodimer complex to the collagen was increased. The binding was almost perfectly inhibited in the coexistence of 10  $\mu$ g/ml of anti-human integrin  $\alpha 2$  antibody (clone P1E6) and anti-human integrin  $\beta 1$  antibody (clone 4B4),

or in the presence of 5 mM EDTA respectively. This result shows that  $\alpha 2 \cdot \lg G$  heavy chain- $\beta 1 \cdot \lg G$  heavy chain chimeric protein heterodimer complex can be bound to a collagen like the integrin  $\alpha 2\beta 1$  existing on the surfaces of cell membranes, and furthermore that the binding is  $\alpha 2\beta 1$ -specific and that the feature of the binding that it depends on cations is retained.

#### Example 17

Acquisition of a peptide capable of being bound to  $\alpha 4 \cdot lgG$  heavy chain chimeric protein heterodimer complex, and evaluation of its inhibitory activity

At first,  $\alpha 4 \cdot \lg G$  heavy chain- $\beta 1 \cdot \lg G$  heavy chain chimeric protein heterodimer complex purified in Example 6, or human  $\lg G$  was prepared at a proper concentration by PBS(-) and was coated on a plastic plate at  $4^{\circ}C$  for 16 hours, being formed as a solid phase on a plastic plate. Then, according to a report (Cott, J. K. and Smoth, G. P., Science, 249, 386-390 (1990)), a phage peptide library in which a random six amino acid residues were cyclyzed by the disulfide bond of cysteine at both the ends was prepared and suspended in 0.1% BSA-containing TBS buffer. The phage peptide library was reacted with human  $\lg G$  at  $30^{\circ}C$  for 3 hours, to absorb phage peptides capable of being bound to  $\lg G$ . Then, the non-absorbed phases were reacted with  $\alpha 4 \cdot \lg G$  heavy chain- $\beta 1 \cdot \lg G$  heavy chain chimeric protein heterodimer complex at  $30^{\circ}C$  for 3 hours, and

the reaction mixture was washed with 0.1% BSA-containing TBS buffer twice to remove the phage peptides incapable of being bound to the heterodimer complex. Only the phage peptides capable of being bound were collected after elution with 0.1M glycine-hydrochloric acid (pH 2.2). After collection, the phage was amplified and the above mentioned binding operation was repeated further twice. The only the phage peptides capable of being bound to the heterodimer complex were selectively concentrated. In the final elution operation, phage peptides capable of being bound to the heterodimer complex were eluted using 10 mM EDTA and 0.1M glycinehydrochloric acid in two steps, and the amino acid sequences of the respective peptides were analyzed. Of them, eight sequences (sequence Nos. 24 to 31) are shown in Table 1. Furthermore, they were examined using the binding assay system of Example 9, and the IC50 values of the four peptide sequences showing binding inhibitory activity are shown in Table 1.

Table 1

Elution condition		Sequence								Sequence No.
	Cys*	lle	Pro	Glu	Leu	lle	Vai	Cys*	1.2	24
EDTA	Cys*	Met	Arg	Tyr	Thr	Ser	Ala	Cys*	2.3	2 5
	Cys*	Glu	Trp	Met	Lys	Arg	Phe	Cys*		2 6
	Cys*	Tyr	Thr	Thr	Arg	Leu	Lys	Cys*		27

Glycine-	Cys*	Leu	Arg	Tyr	Ser	Va!	Pro	Cys*	1.8	28
hydrochloric acid	Cys*	lle	Val	Asn	Arg	Leu	Gly	Cys*		29
	Cys*	Gly	Leu	Gln	Ala	Leu	Pro	Cys*	10	30
	Cys*	Lys	Leu	Lys	Gly	Thr	Met	Cys*		31

Cys\* indicates a disulfide bond.

#### Example 18

Acquisition of a low weight molecular compound capable of inhibiting the binding between the peptide fragment on fibronectin and  $\alpha 4 \cdot \lg G$  heavy chain- $\beta 1 \cdot \lg G$  heavy chain chimeric protein heterodimer complex

Reagents and reported compounds were picked up at random, adjusted to a final concentration of 50 or 100  $\mu g/ml$ , and added to the binding determination system in Example 9. Compounds showing inhibitory activity were obtained. Of the obtained compounds, the binding inhibitory activities of the four compounds of Norethynodrel (Sigma), D-Penicillamine (Aldrich, Weigert, W. M. et al., Angew. Chem. Int. Ed. Eng., 14, 330-336 (1975),  $\gamma$ -2-Naphthyl butyric acid (Fieser, L. F. J. Am. Chem. Soc., 70, 3197-3203 (1948)), 1-Adamantaneacetic acid (Aldrich) were shown in Table 2.

Table 2

Name of compound	Concentration (µg/ml)	Inhibition rate (%)
Norethynodrel	50	2 8
D-Penicillamine	50	51
γ-2-Naphtyl butyric acid	100	37
I-Adamantaneacetic acid	100	65

# Example 19

Preparation of  $\alpha 2 \cdot \lg G$  heavy chain- $\beta 1 \cdot \lg G$  heavy chain chimeric protein heterodimer complex liposome

A liposome was prepared according to the Martin et al.'s method (Martin, F. J. et al., Biochemistry, 20, 4229, (1981)). At first, an activated SH group was introduced into dipalmitoyl phosphatidyl ethanolamine (DPPE, Sigma) using dicrosslinking reagent N-succineimidy! 3-(2pyridyldithio)propionate (SDPD, Sigma), to prepare pyridylthiopropionyl dipalmitoyl phosphatidyl ethanolamine (PDP-DPPE). The PDP-DPPE, dipalmitoyl phosphatidyl choline (DPPC) and cholesterol were mixed, to prepare a lipid film, and it was treated by a sonicator. Then, a filter was used to obtain a liposome uniform in diameter (PDP-DPPE liposome). Then,  $\alpha 2 \cdot \lg G$  heavy chain- $\beta 1 \cdot \lg G$  heavy chain chimeric protein heterodimer complex or human IgG (Cappel) used as a negative control were dissolved in a Hepes buffer (100 mM Hepes, 150 mM NaCl, pH 8.0), and SDPD was added for reaction for 30 minutes. The reaction solution was applied to PD-10 column (Pharmacia), and eluted by 0.1M acetic acid-sodium acetate buffer (pH 5.5). To the eluate, dithiothreitol was added for treatment for 20 minutes, and the mixture was applied to PD-10 column again and eluted by a Hepes buffer (100 mM Hepes, 150 mM NaCl, pH 8.0), to obtain SDPD coupled  $\alpha 2 \cdot \lg G$  heavy chain- $\beta 1 \cdot \lg G$  heavy chain chimeric protein heterodimer complex. The SDPD modified heterodimer complex and the PDP-DPPE liposome were caused to react with each other at room temperature for 24 hours, and the reaction mixture was separated by Sepharose 4B column (Sigma). From the peak fraction,  $\alpha 2 \cdot \lg G$  heavy chain- $\beta 1 \cdot \lg G$  heavy chain chimeric protein heterodimer complex liposome was obtained.

The amount of  $\alpha 2 \cdot \lg G$  heavy chain— $\beta 1 \cdot \lg G$  heavy chain chimeric protein heterodimer complex bound on the liposome was determined by a densitometer (ATTO) after SDS-PAGE/Coummassie staining, and adjusted to final concentration of 1 mg/ml. Example 20

Flow cytometry of  $\alpha 2 \cdot \lg G$  heavy chain- $\beta 1 \cdot \lg G$  heavy chain chimeric protein heterodimer complex liposome

 $\alpha 2 \cdot \text{lgG}$  heavy chain-\$\beta 1 \cdot \text{lgG} heavy chain chimeric protein heterodimer complex liposome was dispersed in 1 mM EDTA-containing PBS(-), and caused to react with anti-human integrin \$\alpha 2\$ antibody (clone P1E6) or anti-human integrin \$\beta 1\$ antibody (clone 4B4) at room temperature for 30 minutes. After completion of reaction, the reaction mixture was centrifuged at 15000 rpm for 10 minutes, being followed by washing with 1 mM EDTA-containing PBS(-) and suspended into the solution again. Into the suspension, FITC labeled anti-murine lgG antibody (Cappel, 10 \$\mu g/ml\$) was added as a secondary antibody, and reacted at room temperature for 30

minutes. After completion of reaction, the reaction mixture was similarly washed by centrifugation, and flow cytometry analysis (ELITE, Coulter) was performed.

.As a result, the positive reactions for both the antibodies were detected, confirming that  $\alpha 2 \cdot \lg G$  heavy chain-  $\beta 1 \cdot \lg G$  heavy chain chimeric protein heterodimer complex was bound on the liposome.

Example 21

Binding activity of  $\alpha 2$  -  $\lg G$  heavy chain- $\beta 1$  -  $\lg G$  heavy chain chimeric protein heterodimer complex to a collagen

A collagen (Cell Matrix Typel, 3 mg/ml) was diluted by 0.02M acetic acid solution, and the solution was put in an immunoplate by 100  $\mu$ l/well, being kept at 4°C for 16 hours. Then, the collagen solution was removed by suction, and the plate was washed with PBS(-) twice for neutralization, and heat-denaturated 1% BSA-PBS solution was put in the plate by 300  $\mu$ l/well for blocking at room temperature for 3 hours. After completion of blocking, the plate was rinsed with PBS(-) twice, to prepare a collagen coated plate.

Normal human plasma (George King) and von Willebrand's factor deficient (severe) plasma (George Kind) were treated with anti-human IgG antibody and protein A, and dialyzed against PBS(-) for 24 hours, to remove the contained sodium citrate. In order that the Ca ion and Mg ion concentration might be a physiological concentration in the blood when used,

CaCl<sub>2</sub> and MgCl<sub>2</sub> were added to achieve final concentrations of 1.2 mM and 0.2 mM respectively. Into the normal human plasma and von Willebrand's factor deficient plasma adjusted in cation concentration,  $\alpha 2 \cdot \lg G$  heavy chain  $-\beta 1 \cdot \lg G$  heavy chain chimeric protein heterodimer complex liposome or human IgG liposome was suspended to achieve protein concentrations of 1 to 100 ng/ml. Any of the suspensions was put in the collagen coated plate by 100  $\mu$ l/well. The plate was shaken by a plate shaker at 100 rpm, for reaction at room temperature for 15 minutes. After completion of reaction, the non-bound liposome was removed by washing with a PB solution (1.2 mM CaCl<sub>2</sub>, 0.2 mM MgCl<sub>2</sub>, 1% BSA-containing PBS, pH 7.4), and the bound liposome was immobilized by 1% glutaraldehyde-PBS at room temperature for 30 minutes. After completion of immobilization, a heatdenaturated BSA-PBS solution was used for blocking at room temperature for 1 hour. Then, as described in Example 16, it was caused to react with biotin labeled human lgG antibody used as a primary antibody and avidin labeled horseradish peroxidase used as a secondary antibody, and washed with a TBS buffer. Into it, orthophenylenediamine was added as a substrate for color development, and the absorbance at 490 nm were measured. To examine the effect of 5 mM EDTA, antiintegrin  $\alpha$ 2 antibody (clone P1E6, 10  $\mu$ g/ml) and anti-integrin  $\beta$ 1 antibody (clone 4B4, 10  $\mu$ g/ml), it was caused to react with the liposome suspension at room temperature for 15 minutes

before reaction with the collagen.

The results are shown in Figs. 5 and 6. In the normal human plasma, the human lgG liposome as a negative control was not found to be bound to the collagen, but the binding of  $\alpha 2$  -IgG heavy chain-\$1 · IgG heavy chain chimeric protein heterodimer complex liposome to the collagen was increased with the concentration dependent on manner. Also when the von Willebrand's factor deficient plasma was used, equivalent binding was detected. Furthermore, the binding to the collagen observed when 30 ng/ml of  $\alpha 2 \cdot \text{lgG}$  heavy chain- $\beta 1 \cdot \text{lgG}$ heavy chain chimeric protein heterodimer complex liposome was added to the normal plasma was completely inhibited by adding EDTA as a cation chelating agent or the antibodies. results show that in plasma with a physiological cation concentrtion,  $\alpha 2 \cdot \text{lgG}$  heavy chain- $\beta 1 \cdot \text{lgG}$  heavy chain chimeric protein heterodimer complex liposome is bound to a collagen like platelets, and strongly suggest that it can be a substitute of adhesive platelets, and can be a reagent for monitoring the collagen exposed region. Furthermore, it is indicated that since equivalent binding activity was shown also in von Willebrand's factor defincient plasma, the liposome can also be used in the plasma with coagulation abnormality such as von Willebrand's disease.

Example 22

Analysis of collagen covering state by  $\alpha 2 \cdot lgG$  heavy chain- $\beta$ 

1 - IgG heavy chain chimeric protein heterodimer complex liposome

Five microliters of a collagen solution was spotted at the center of each of the wells of a Lab-Tek chamber slide (Intermed, 8-well type, plastic) and allowed to stand for 16 hours, then washed and treated for blocking. Then, a suspension in which  $\alpha 2 \cdot \lg G$  heavy chain- $\beta 1 \cdot \lg G$  heavy chain chimeric protein heterodimer complex liposome was suspended in normal human plasma to achieve a protein concentration of 30 ng/ml as described in Example 21 was put in the slide by 200  $\mu$ l/well, for reaction under the same conditions. After completion of reaction, the non-bound liposome was removed by washing with a PB buffer, and the retained was immobilized and treated for blocking. Then, it was bound to biotin labeled anti-human IgG antibody as a primary antibody and with avidin labeled horseradish peroxidase as a secondary antibody, and was washed with a TBS buffer. After completion of washing, diaminobenzidine was added for staining, to observe the covering state of the  $\alpha 2 \cdot \text{lgG}$  heavy chain- $\beta 1 \cdot \text{lgG}$  heavy chain chimeric protein heterodimer complex liposome bound on the collagen.

With the human IgG liposome, the collagen coated portion was not stained, but with  $\alpha 2 \cdot \text{lgG}$  heavy chain- $\beta 1 \cdot \text{lgG}$  heavy chain chimeric protein heterodimer complex liposome, the collagen coated portion was entirely stained. Therefore,

since the  $\alpha 2 \cdot \lg G$  heavy chain- $\beta 1 \cdot \lg G$  heavy chain chimeric protein heterodimer complex liposome covered the collagen coated portion only, it was strongly suggested that the liposome could be a substitute of adhesive platelets. Industrial Availability:

The present invention provides integrin-immunoglobulin chimeric protein heterodimer complexes in which the  $\alpha$  chain and the  $\beta$  chain of an integrin are stably associated. The obtained integrin-immunoglobulin chimeric protein heterodimer complexes can be directly used as drugs, and can also be used for determining the binding between an integrin and a ligand, and searching for a substance capable of being bound to an integrin and a substance inhibiting the binding between an integrin and a ligand. They can also be used as diagnostic reagents.

Furthermore, among the heterodimer comlexes, especially integrin  $\alpha 2\beta 1$ -immunoglobulin chimeric protein heterodimer complex can be used as a substitute of platelets. Furthermore, integrin  $\alpha 2\beta 1$ -immunoglobulin chimeric protein heterodimer complex can be used as a therapeutic or preventive agent for bleeding tendency involved in thrombocytopenia, platelet function abnormality, etc. Furthermore, it can also be used as a reagent for monitoring the exposed region of an extracellular matrix and for the targeting therapy.

## Sequence Table

Sequence No. 1

Length of sequence: 4228

Type of sequence: Nucleic acid

Sequence

ATG	TTC	ССС	ACC	GAG	AGC	GCA	TGG	CTT	GGG	AAG	CGA	GGC	GCG	AAC	CCG	48
Жet	Phe	Pro	Thr	Glu	Ser	Ala	Trp	Leu	Gly	Lys	Arg	Gly	Ala	Asn	Pro	
				-35					-30					-25		
GGC	ССС	GAA	GCT	GCA	CTC	CGG	GAG	ACG	GTG	ATG	CTG	TTG	CTG-	-TGC	CTG	96
Gly	Pro	Glu	Ala	Ala	Leu	Arg	Głu	Thr	Val	Met	Leu	Leu	Leu	Cys	Leu	
			-20					-15					-10			
GGG	GTC	CCG	ACC	GGC	AGG	CCT	TAC	AAC	GTG	GAC	ACT	GAG	AGC	GCG	CTG	144
Gĺy	Val	Pro	Thr	Gly	Arg	Pro	Tyr	Asn	Val	Asp	Thr	Glu	Ser	Ala	Leu	
		- 5					1				5					
CTT	TAC	CAG	GGC	ССС	CAC	AAC	ACG	CTG	TTC	GGC	TAC	TCG	GTC	GTG	CTG	192
Leu	Tyr	Gln	Gly	Pro	His	Asn	Thr	Leu	Phe	Gly	Tyr	Ser	Val	Val	Leu	
10					15					20					25	
CAC	AGC	CAC	GGG	GCG	AAC	CGA	TGG	CTC	CTA	GTG	GGT	GCG	CCC	ACT	GCC	240
His	Ser	His	Gly	Ala	Asn	Arg	Trp	Leu	Leu	Val	Gly	Ala	Pro	Thr	Ala	
				30			-		35					40		
AAC	TGG	CTC	GCC	AAC	GCT	TCA	GTG	ATC	AAT	ccc	GGG	GCG	ATT	TAC	AGA	288
Asn	Trp	Leu	Ala	Asn	Ala	Ser	Val	Ile	Asn	Pro	Gly	Ala	Ile	Tyr	Arg	
			45					50					55			
TGC	AGG	ATC	GGA	AAG	AAT	ССС	GGC	CAG	ACG	TGC	G A A	CAG	СТС	CAG	CTG	336
Cys	Arg	lle	Gly	Lys	Asn	Pro	Gly	Gln	Thr	Cys	Glu	Gln	Leu	Gln	Leu	
		60					6 5					70				
GGT	, VCC	сст	' ለለፕ	. GCV	GAA	сст	тст	, CCV	λλG	λСΊ	TGT	TTG	GAA	GAG	λGλ	384
Gly	Ser	Pro	Asr	Gly	Gli	Pro	Cys	Gly	Lys	Th r	Cys	Leu	Glu	Glu	λrg	
	75	, )				80	<b>)</b>				85	i				

GVC	λλΤ	CAG	TGG	TTG	GGG	GTC	ACA	СТТ	TCC	AGA	CAG	CCY	GGA	GAA	TAK	432
Asp	Asn	Gln	Trp	Leu	Gly	Val	Thr	Leu	Ser	Arg	Gln	Pro	Gly	Glu	λsπ	
90					95					100					105	
GGA	TCC	ATC	GTG	ACT	TGT	GGG	СЛТ	A G A	TGG	A A A	T A A	ATA	TTT	TAC	ATA	480
Gly	Ser	Ile	Val	Thr	Cys	Gly	His	Arg	Trp	Lys	Asn	Ile	Phe	Tyr	lle	
				110					115					120		
AAG	AAT	GAA	AAT	AAG	СТС	CCC	ACT	GGT	GGT	TGC	TAT	GGA	GTG	ССС	CCT	528
Lys	Asn	Glu	Asn	Lys	Leu	Pro	Thr	Gly	Gly	Cys	Tyr	Gly	Val	Pro	Pro	
			125					130					135			
GAT	TTA	CGA	ACA	GAA	CTG	AGT	AAA	AGA	ATA	GCT	CCG	TGT	TAT	CAA	GAT	576
Asp	Leu	Arg	Thr	Glu	Leu	Ser	Lys	Arg	Ile	Ala	Pro	Cys	Tyr	Gln	Asp	
		140					145					150				
TAT	GTG	A A A	A A A	TTT	GGA	GAA	AAT	TTT	GCA	TCA	TGT	CAA	GCT	GGA	ATA	624
Tyr	Val	Lys	Lys	Phe	Gly	Glu	Asn	Phe	Ala	Ser	Cys	Gln	Ala	Gly	Ile	
	155					160					165					
TCC	AGT	TTT	TAC	ACA	AAG	GAT	TTA	ATT	GTG	ATG	GGG	GCC	CCA	GGA	TCA	672
Ser	Ser	Phe	Tyr	Thr	Lys	Asp	Leu	Ile	Val	Met	Gly	Ala	Pro	Gly	Ser	
170					175					180					185	
TCT	TAC	TGG	ACT	GGC	TCT	CTT	TTT	GTC	TAC	AAT	ATA	ACT	ACA	AAT	AAA,	720
Ser	Tyr	Trp	Thr	Gly	Ser	Leu	Phe	Val	Tyr	Asn	Ile	Thr	Thr	Asn	Lys	
				190					195					200		
TAC	AAG	GCT	TTT	TTA	GAC	AAA	CAA	AAT	CAA	GTA	AAA	ŢŢŢ	GGA	AGT	TAT	768
Tyr	Lys	Ala	Phe	Leu	Asp	Lys	G1n	Asn	Gln	Val	Lys	Phe	Gly	Ser	Tyr	
			205					210					215			
TTA	GGA	TAT	TCA	GTC	GGA	GCT	GGT	CAT	TTT	CGG	AGC	CAG	CAT	ACT	ACC	816
Leu	Gly	Tyr	Ser	Val	Gly	Ala	Gly	His	Phe	Arg	Ser	Gln	His	Thr	Thr	
		220					225					230				
GAA	GTA	GTC	GGA	GGA	GCT	CCT	CAA	CAT	CAC	CAG	ATT	GGT	λλC	GCA	TAT	864
Glu	Val	Val	Gly	Gly	λla	Pro	Gln	llis	Glu	Gln	lle	Gly	Lys	λla	Туг	
	235					240					245					

ATA	TTC	AGC	TTK	GAT	GAA	hhh	GAA	CTA	AAT	ATC	TTA	CAT	CAA	ATG	AAA	912
lle	Phe	Ser	lle	Asp	Glu	Lys	Glu	Leu	Asn	lle	Leu	His	Glu	Меt	Lys	
250					255					260					265	
GGT	ΛAA	AΛG	CTT	GGA	TCG	TAC	TTT	GGA	GCT	TCT	GTC	TGT	GCT	GTG	GAC	960
Gly	Lys	Lys	Leu	Gly	Ser	Туr	Phe	Gly	Ala	Ser	Val	Cys	Ala	Val	Asp	
				270					275					280		
CTC	AAT	GCA	GAT	GGC	TTC	TCA	GAT	CTG	CTC	GTG	GGA	GCA	CCC	АTС	CAG	.1008
Leu	Asn	Ala	Asp	Gly	Phe	Ser	Asp	Leu	Leu	Val	Gly	Ala	Pro	Меt	Gln	
			285			•		290					295			
AGC	ACC	ATC	AGA	GAG	GAA	GGA	AGA	GTG	TTT	GTG	TAC	ATC	AAC.	TCT	GGC	1056
Ser	Thr	Ile	Arg	Glu	Glu	Gly	Arg	Val	Phe	Vаl	Tyr	Ile	Asn	Ser	Gly	
		300					305					310			,	
TCG	GGA	GCA	GTA	ATG	AAT	GCA	ATG	GAA	ACA	AAC	CTC	GTT	GGA	AGT	GAC	1104
Ser	Gly	Ala	Val	Меt	Asn	Ala	Met	Glu	Thr	Asn	Leu	Val	Gly	Ser	Asp	
	315					320					325				•	
AAA	TAT	GCT	GCA	AGA	TTT	GGG	GAA	TCT	ATA	GTT	AAT	CTT	GGC	GAC	ATT	1152
Lys	Tyr	Ala	Ala	Arg	Phe	Gly	Glu	Ser	Ile	Val	Asn	Leu	Gly	Asp	Ile	
330					335					340					345	
GAC	AAT	GAT	GGC	TTT	GAA	GAT	GTT	GCT	ATC	GGA	GCT	CCA	CAA	GAA	GAT	1200
Asp	Asn	Asp	Gly	Phe	Glu	Asp	Val	Ala	Ile	Gly	Ala	Pro	Gln	Glu	Asp	
				350					355					360		
GAC	TTG	CAA	GGT	GCT	ATT	TAT	ATT	TAC	AAT	GGC	CGT	GCA	GAT	GGG	ATC	1248
Asp	Leu	Gln	Gly	Ala	Ile	Tyr	[le	Tyr	Asn	Gly	Arg	Ala	Asp	Gly	lle	
			365					370					375			
TCG	TCA	ACC	TTC	TCA	CAG	AGA	ATT	GAA	GGA	CTT	CAG	ATC	AGC	AAA	TCG	1296
Ser	Ser	Thr	Phe	Ser	Gln	λrg	lle	Glu	Gly	Leu	Gln	lle	Ser	Lys	Ser	
		380					385					390				
TTA	ΛGT	ΛTC	TTT	$GG\Lambda$	CAG	тст	ATA	ТСЛ	CGA	CAA	ATT	GAT	GCA	GAT	TAA	1344
Leu	Ser	Met	Phe	Gly	Gln	Ser	He	Ser	Gly	Gln	He	λερ	Лlа	λsp	λsn	
	395					400					405					

AAT	GGC	TAT	GTA	GAT	GTA	GCA	GTT	GGT	GCT	TTT	CGG	TCT	GAT	TCT	GCT	1392
Asn	Gly	Туr	Va l	Asp	Val	Ala	Val	Gly	Ala	Phe	Arg	Ser	λsρ	Ser	Ala	
410					415					420					425	
GTC	TTG	СТА	AGG	ACA	A G A	CCT	GTA	GTA	ATT	GTT	GAC	GCT	TCT	TTA	AGC	1440
Val	Leu	Leu	Arg	Thr	Arg	Pro	۷al	Val	ſle	Val	Asp	Ala	Ser	Leu	Ser	
				430					435					440		
CAC	CCT	GAG	TCA	GTA	AAT	AGA	ACG	AAA	TTT	GAC	TGT	GTT	GAA	AAT	GGA	1488
His	Pro	Glu	Ser	Val	Asn	Arg	Thr	Lys	Phe	Asp	Cys	Val	Glu	Asn	Gly	
			445					450					455			
TGG	CCT	TCT	GTG	TGC	ATA	GAT	CTA	A C A	CTT	TGT	TTC	TCA	TAT	AAG	GGC	1536
Trp	Pro	Ser	Val	Cys	Ile	Asp	Leu	Thr	Leu	Cys	Phe	Ser	Tyr	Lys	Gly	
		460					465					470				
A A G	GAA	GTT	CCA	GGT	TAC	ATT	GTT	TTG	TTT	TAT	AAC	ATG	AGT	TTG	GAT	1584
Lys	Glu	Yal	Pro	Gly	Tyr	Ile	Val	Leu	Phe	Tyr	Asn	Met	Ser	Leu	Asp	
	475					480					485					
GTG	AAC	AGA	AAG	GCA	GAG	TCT	CCA	CCA	AGA	TTC	TAT	TTC	TCT	TCT	AAT	1632
Val	Asn	Arg	Lys	Ala	Glu	Ser	Pro	Pro	Arg	Phe	Tyr	Phe	Ser	Ser	Asn	
490					495					500					505	
GGA	ACT	TCT	GAC	GTG	ATT	ACA	GGA	AGC	ATA	CAG	GTG	TCC	AGC	AGA	GAA	1680
Gly	Thr	Ser	Asp	Val	Ile	Thr	G1y	Ser	Ile	Gln	Уаl	Ser	Ser	Arg	Glu	
				510					515					520		
GCT	AAC	TGT	AGA	ACA	CAT	CAA	GCA	TTT	ATG	CGG	AAA	GAT	GTG	CGG	GAC	1728
Ala	Asn	Cys	Arg	Thr	His	Gln	Ala	Phe	Met	Arg	Lys	Asp	Yal	Arg	Asp	
			525					530					535			
ATC	CTC	ACC	CCA	ATT	CAG	ATT	GAA	GCT	GCT	TAC	CAC	CTT	GGT	CCT	CAT	1776
lle	Leu	Thr	Pro	lle	Gln	lle	Glu	Ala	Ala	Ţуr	His	Leu	Gly	Pro	His	
		540					545					550				
GTC	ATC	λGT	AAA	CGA	λGT	λСΛ	GAG	GAA	TTC	CCA	CCA	CTT	CAG	CCA	ΤΤκ	1824
Val	lle	Ser	Lys	Arg	Ser	Thr	Glu	Glu	Phe	Pro	Pro	Leu	Gln	Pro	He	
	555					560					565					

CII	CAG	CAG	AAG	AAA	GAA	AAA	GAC	ATA	λTG	AAA	AAA	ACA	AIA	AAC	111	1872
Leu	Gln	Gln	Lys	Lys	Glu	Lys	Asp	lle	Меt	Lys	Lys	Thr	lle	λsn	Phe	
570					575					580					585	
GCA	AGG	TTT	TGT	GCC	CAT	GAA	AAT	TGT	TCT	GCT	GAT	ATT	CAG	GTT	TCT	1920
Ala	λrg	Phe	Cys	Ala	His	Glu	Asn	Cys	Ser	Ala	Asp	Leu	Gln	Val	Ser	
	,			590					595					600		
GCA	AAG	ATT	GGG	TTT	TTG	AAG	CCC	CAT	GAA	AAT	AAA	ACA	TAT	CTT	GCT	1968
Ala	Lys	[le	Gly	Phe	Leu	Lys	Pro	His	Glu	Asn	Lys	Thr	Tyr	Leu	Ala	
			605					610					615			
GTT	GGG	AGT	ATG	AAG	ACA	TTG	ATG	TTG	AAT	GTG	TCC	TTG	TTT	AAT	GCT	2016
Val	Gly	Ser	Жet	Lys	Thr	Leu	Жet	Leu	Asn	Yal	Ser	Leu	Phe	Asn	Ala	
		620					625					630				
GGA	GAT	GAT	GCA	TAT	GAA	ACG	ACT	CTA	CAT	GTC	AAA	CTA	CCC	GTG	GGT	2064
Gly	Asp	Asp	Ala	Tyr	Glu	Thr	Thr	Leu	His	Yal	Lys	Leu	Pro	Yal	Gly	
	635					640					645					
CTT	TAT	TTC	ATT	AAG	ATT	TTA	GAG	CTG	GAA	GAG	AAG	CAA	ATA	AAC	TGT	2112
Leu	Tyr	Phe	Ile	Lys	Ile	Leu	Glu	Leu	Glu	Glu	Lys	Gln	Ile	Asn	Cys	
650					655				•	660					665	
GAA	GTC	ACA	GAT	AAC	TCT	GGC	GTG	GTA	CAA	CTT	GAC	TGC	AGT	TTA	GGC	2160
Glu	Yal	Thr	Аsр	Asn	Ser	Gly	Val	Val	Gln	Leu	Asp	Cys	Ser	lle	Gly	
				670					675					680		
TAT	ATA	TAT	GTA	GAT	CAT	CTC	TCA	AGG	ATA	GAT	ATT	AGC	TTT	CTC	CTG	2208
Tyr	lle	Tyr	Ya1	Asp	His	Leu	Ser	Arg	Ile	Asp	Ile	Ser	Phe	Leu	Leu	
			685					690					695			
GAT	GTG	AGC	TCA	CTC	AGC	AGA	GCG	GAA	GAG	GAC	CTC	AGT	ATC	ACA	GTG	2256
Аsр	Yal	Ser	Ser	Leu	Ser	Arg	Ala	Glu	Glu	Asp	Leu	Ser	lle	Thr	Val	
		700					705					710				
CAT	GCT	VCC	TGT	GAA	TAK	GAA	GΛG	$GA\Lambda$	ATG	GAC	ለለፐ	СТЛ	AAG	CVC	AGC	2304
llis	λla	Thr	Cys	Glu	Лsп	Glu	Glu	Glu	Met	λsρ	Asn	Leu	Lys	llis	Ser	
	715					720					725					

			_				m.m		<b></b>	CLC	ሶጥጥ	110	CTC	1СТ	СТТ	2352
						CCT	-									2002
Arg	Val	Thr	Val	Ala	lle	Pro	Leu	Lys	Туr	Glu	Val	Lys	Leu	Thr		
730					735					740					745	
CAT	GGG	TTT	GTA	AAC	CCA	ACT	TCA	TTT	GTG	TAT	GGA	TCA	AAT	GAT	GAA	2400
His	Gly	Phe	Val	Asn	Pro	Thr	Ser	Phe	Val	Туr	Gly	Ser	Asn	Asp	Glu	
				750	-		-		755					760		
AAT	GAG	CCT	GAA	ACG	TGC	ATG	GTG	GAG	AAA	ATG	AAC	TTA	ACT	TTC	CAT	2448
Asn	Glu	Pro	Glu	Thr	Cys	Met	Val	Glu	Lys	Met	Asn	Leu	Thr	Phe	His	
			765					770					775			
GTT	ATC	AAC	ACT	GGC	AAT	AGT	ATG	GCT	CCC	AAT	GTT	AGT	GTG.	G A A	ATA	2496
Val	Ile	Asn	Thr	Gly	Asn	Ser	Met	Ala	Pro	Asn	Yal	Ser	Val	Glu	.Ile	
		780					785					790				
ATG	GTA	CCA	AAT	TCT	TTT	AGC	CCC	CAA	ACT	GAT	AAG	CTG	TTC	AAC	ATT.	2588
						Ser										
	795					800					805					
TTG	GAT	GTC	CAG	ACT	ACT	ACT	GGA	G A A	TGC	CAC	TTT	GAA	AAT	TAT	CAA	2592
						Thr										-
810					815				•	820					825	
AGA	GTG	TGT	GCA	TTA	GAG	CAG	CAA	AAG	AGT	GCA	ATG	CAG	ACC	TTG	AAA	2640
						Gln										
				830					835					840		•
GGC	ATA	GTC	CGG	TTC	TTG	TCC	A A G	ACT	GAT	AAG	AGG	CTA	TTG	TAC	TGÇ	2688
															Cys	
			845					850					855			
ATA	AAA	GCT	' GAT	CCA	CAT	TGT	TTA	AAT	TTC	TTG	TGT	AAT	TTT	GGG	AAA	2736
															Lys	
		860					865					870				
ATO	GAA				G A A	GCC	AG1	GT1	CAT	. 410	CAA	CTG	G A A	GGG	CGG	2784
															/ Arg	
	875		. ,	•		880					885					
	5,0	•														

CCA TCC ATT TTA GAA ATG GAT GAG ACT TCA GCA CTC AAG TTT GAA ATA	2832
Pro Ser Ile Leu Glu Met Asp Glu Thr Ser Ala Leu Lys Phe Glu Ile	
890 895 900 905	
AGA GCA ACA GGT TTT CCA GAG CCA AAT CCA AGA GTA ATT GAA CTA AAC	2880
Arg Ala Thr Gly Phe Pro Glu Pro Asn Pro Arg Val [le Glu Leu Asn	
910 915 920	
AAG GAT GAG AAT GTT GCG CAT GTT CTA CTG GAA GGA CTA CAT CAA	2928
Lys Asp Glu Asn Val Ala His Val Leu Leu Glu Gly Leu His His Gln.	
925 930 935	
AGA CCC AAA CGT TAT TTC ACG GAT CCC GAG CTGCTGGAAG CAGGCTCAGC	2978
Arg Pro Lys Arg Tyr Phe Thr Asp Pro Glu	
940 945	
GCTCCTGCCT GGACGCATCC CGGCTATGCA GCCCCAGTCC AGGGCAGCAA GGCAGGCCCC	3038
GTCTGCCTCT TCACCCGGAG CCTCTGCCCG CCCCACTCAT GCTCAGGGAG AGGGTCTTCT	3098
GGCTTTTTCC CAGGCTCTGG GCAGGCACAG GCTAGGTGCC CCTAACCCAG GCCCTGCACA	3158
CAAAGGGGCA GGTGCTGGGC TCAGACCTGC CAAGAGCCAT ATCCGGGAGG ACCCTGCCCC	3218
TGACCTAAGC CCACCCCAAA GGCCAAACTC TCCACTCCCT CAGCTCGGAC ACCTTCTCTC	3278
CTCCCAGATT CCAGTAACTC CCAATCTTCT CTCTGCA GAG CCC AAA TCT TGT GAC	3333
Glu Pro Lys Ser Cys Asp	
950	
AAA ACT CAC ACA TGC CCA CCG TGC CCA GGTAAGCCAG CCCAGGCCTC	3380
Lys Thr His Thr Cys Pro Pro Cys Pro	·
955 960	
GCCCTCCAGC TCAAGGCGGG ACAGGTGCCC TAGAGTAGCC TGCATCCAGG GACAGGCCCC	3440
AGCCGGGTGC TGACACGTCC ACCTCCATCT CTTCCTCA GCA CCT GAA CTC CTG	3493
Ala Pro Glu Leu Leu	
965	
GGG GGA CCG TCA GTC TTC CTC TTC CCC CCA AAA CCC AAG GAC ACC CTC	3541
Gly Gly Pro Ser Val Phe Leu Phe Pro Pro Lys Pro Lys Asp Thr Leu	
970 975 980	

AT G	λTC	TCC	CGG	ACC	CCT	GAG	GTC	ACA	TGC	GTG	GTG	GTG	GAC	GTG	ЛGС	3589
let	[le	Ser	Arg	Thr	Pro	Glu	Val	Thr	Cys	Val	Val	Val	λsp	Val	Ser	
	985					990					995					
CAC	GAA	GAC	CCT	GAG	GTC	A A G	TTC	AAC	TGG	TAC	GTG	GAC	GGC	GTG	GAG	3637
His	Glu	Asp	Pro	Glu	Val	Lys	Phe	Asn	Trp	Tyr	Val	Asp	Gly	Val	Glu	
1000	١			1	005					1010					1015	
GTG	CAT	AAT	GCC	AAG	ACA	AAG	CCG	CGG	GAG	GAG	CAG	TAC	AAC	AGC	ACG	3685
Val	His	Asn	Ala	Lys	Thr	Lys	Pro	Arg	Glu	Glu	Gln	Tyr	Asn	Ser	Thr	
			]	1020					1025					1030		
TAC	CGG	GTG	GTC	AGC	GTC	CTC	ACC	GTC	CTG	CAC	CAG	GAC	TGG	CTG	AAT	3733
Tyr	Arg	Val	Yal	Ser	Val	Leu	Thr	Val	Leu	His	Gln	Asp	Trp	Leu	Asn	
			1035					1040					1045		•	
GGC	AAG	GAG	TAC	AAG	TGC	AAG	GTC	TCC	AAC	AAA	GCC	CTC	CCA	GCC	CCC	3781
Gly	Lys	Glu	Tyr	Lys	Cys	Lys	Val	Ser	Asn	Lys	Ala	Leu	Pro	Ala	Pro	
		1050					1055					1060				
ATC	GAG	AAA	ACC	ATC	TCC	AAA	GCC	AAA	GGT	GGGA	CCC	GTGG	GGTG	CG		3828
lle	Glu	Lys	Thr	Ile	Ser	Lys	Ala	Lys								
	1065					1070										
															CTGTA(	
CAA	ССТС	TGT	CCTA													3937
				G	ly G	ln P	ro A	rg G	lu P	ro G	In Y	'al T	'yr T	hr L	eu	
							75					080				
															TGC	3985
Pro	Pro	Ser	Arg	, Asp	Glu	Leu	Thr	- Lys	. Asr	ı Glı			Lei	ı Thi	Cys	
	1085					1090					1095					
															G AGC	4033
Lei	ı Val	Lys	s Gly	7 Phe	e Tyr	Pro	Se i	r Ası	o II			l Gli	ı Trş	p Gli	ı Ser	
110	0 (				1105	j				111	0				1115	

AAT	GGG	CAG	CCG	GAG	AAC	AAC	TAC	AAG	ACC	ACC	CCT	CCC	GTG	CTG	GΛT	4081
λsπ	Gly	Gln	Pro	Glu	Asn	Asn	Туr	Ĺуs	Thr	Thr	Pro	Pro	Val	Leu	Asp	
				1120					1125					1130		
TCC	GAC	GGC	TCC	TTC	TTC	CTC	TAC	AGC	AAG	CTC	ACC	GTG	GAC	AAG	AGC	4129
Ser	Asp	Gly	Ser	Phe	Phe	Leu	Tyr	Ser	Lys	Leu	Thr	Val	Asp	Lys	Ser	
			1135					1140					1145			
AGG	TGG	CAG	CAG	GGG	AAC	GTC	TTC	TCA	TGC	TCC	GTG	ATG	CAT	GAG	GCT	4177
Arg	Trp	Gln	Gln	Gly	Asn	Val	Phe	Ser	Cys	Ser	Val	Met	His	Glu	Ala	
		1150				1	155				]	160				
CTG	CAC	AAC	CAC	TAC	ACG	CAG	AAG	AGC	CTC	TCC	CTG	TCT	CCG-	GGT	AAA	4225
Leu	His	Asn	His	Tyr	Thr	Gln	Lys	Ser	Leu	Ser	Leu	Ser	Pro	Gly	Lys	
]	1165				1	.170				1	175					
TGA																4228
Sequ	.enc	e No	. 2													
Leng	th o	of s	equ	ence	e: 3	463										
Гуре	of	seq	uen	ce:	Nuc	leid	c ac	id								
Sequ	.enc	9					,									
ATG	AAT	TTA	CAA	CCA	ATT	TTC	TGG	ATT	GGA	CTG	ATC	AGT	TCA	GTT	TGC	48
Met	Asn	Leu	Gln	Pro	Ile	Phe	Trp	Ile	Gly	Leu	Ile	Ser	Ser	Val	Cys	
-20					-15			•		-10					- 5	
TGT	GTG	TTT	GCT	CAA	ACA	GAT	GAA	AAT	AGA	TGT	TTA	AAA	GCA	AAT	GCC	96
Cys	Уal	Phe	Ala	Gln	Thr	Asp	Glu	Asn	Arg	Cys	Leu	Lys	Ala	Asn	Ala	
				I				5					10		•	
AAA	TCA	TGT	GGA	GAA	TGT	ATA	CAA	GCA	GGG	CCA	AAT	TGT	GGG	TGG	TGC	144
Lys	Ser	Cys	Gly	Glu	Cys	lle	Gln	Ala	Gly	Pro	Asn	Cys	Gly	Trp	Cys	
		15					20					25				
УÇУ	λλΤ	TCA	γСΥ	TTT	ATT	CAG	Gλλ	$GG\Lambda$	ATG	CCT	ACT	TCT	GCV	CGA	TCT	192
Thr	Λsn	Ser	Thr	Phe	Leu	Gln	Glu	Gly	Met	Pro	Thr	Ser	Ala	Arg	Cys	
	30					35					40			,		

GAT	GAT	ATT	GAA	GCC	TTA	A A A	ΑΛG	AAG	GGT	TCC	CCT	CCA	GAT	GAC	ATA	240
Asp	Asp	Leu	Glu	Ala	Leu	Lys	Lys	Lys	Gly	Cys	Pro	Pro	Лsр	Аsр	lle	
45					50					55					60	
G A A	AAT	CCC	A G A	GGC	TCC	AAA	GAT	ATA	AAG	AAA	ААТ	AAA	AAT	GTA	ACC	288
Glu	Asn	Pro	Arg	Gly	Ser	Lys	Asp	lle	Lys	Lys	Asn	Ĺys	Asn	Val	Thr	
				65					70					75		
AAC	CGT	AGC	AAA	GGA	ACA	GCA	GAG	AAG	стс	AAG	CCA	GAG	GAT	ATT	CAT	336
Asn	Arg	Ser	Lys	Gly	Thr	Ala	Glu	Lys	Leu	Lys	Pro	Glu	Asp	Ile	His	
			80					85				•	90			
CAG	ATC	CAA	CCA	CAG	CAG	TTG	GTT	TTG	CGA	TTA	AGA	TCA	GGĢ	GAG	CCA	384
Gln	Ile	Gln	Pro	Gln	Gln	Leu	Val	Leu	Arg	Leu	Arg	Ser	Gly	Glu	Pro	
		95					100					105				
CAG	ACA	TTT	ACA	TTA	AAA	TTC	AAG	AGA	GCT	GAA	GAC	TAT	ccc	ATT	GAC	432
Gln	Thr	Phe	Thr	Leu	Lys	Phe	Lys	Arg	Ala	Glu	Asp	Tyr	Pro	Ile	Asp	
	110					115					120					
CTC	TAC	TAC	CTT	ATG	GAC	CTG	TCT	TAT	TCA	ATG	AAA	GAC	GAT	TTG	GAG	480
Leu	Tyr	Tyr	Leu	Met	Asp	Leu	Ser	Tyr	Ser	Met	Lys	Asp	Asp	Leu	Glu	
125					130				•	135					140	
AAT	GTA	AAA	AGT	CTT	GGA	ACA	GAT	CTG	ATG	AAT	GAA	ATG	AGG	AGG	ATT	528
Asn	Val	Lys	Ser	Leu	Gly	Thr	Asp	Leu	Met	Asn	Glu	Мet	Arg	Arg	Ile.	
				145					150					155		
ACT	TCG	GAC	TTC	AGA	ATT	GGA	TTT	GGC	TCA	TTT	GTG	GAA	AAG	ACT	GTG	576
Thr	Ser	Asp	Phe	Arg	Ile	Gly	Phe	Gly	Ser	Phe	Val	Glu	Lys	Thr	Val	
			160					165					170			
ATG	CCT	TAC	ATT	AGC	ACA	ACA	CCA	GCT	AAG	CTC	AGG	AAC	CCT	TGC	ACA	624
Met	Pro	Tyr	lle	Ser	Thr	Thr	Pro	Ala	Lys	Leu	Arg	Asn	Pro	Cys	Thr	
		175					180					185				
AGT	GAA	CAG	AAC	TGC	VCC	YCC	CCV	TTT	AGC	TAC	444	λλΤ	GTG	CTC	ΛGT	672
Ser	Glu	Gln	λsn	Cys	Thr	Thr	Pro	Phe	Ser	Туr	Lys	λsn	Val	Leu	Ser	
	190					195					200					

CTT	ACT	ΛAT	A A A	GGA	G A A	GTA	TTT	AAT	GAA	CTT	GTT	GGA	h h h	CAG	CGC	720
Leu	Thr	Asn	Lys	Gly	Glu	Va l	P'ne	Asn	Glu	Leu	Val	Gly	Lys	Gln	λrg	
205					210					215					220	
ATA	TCT	GGA	AAT	TTG	GAT	TCT	CCA.	G A A	GGT	GGT	TTC	GAT	GCC	ATC	ATG	768
lle	Ser	Gly	Asn	Leu	Аsр	Ser	Pro	Glu	Gly	Gly	Phe	Asp	Ala	[le	Иet	
				225					230					235		
CAA	GTT	GCA	GTT	TGT	GGA	TCA	CTG	ATT	GGC	TGG	AGG	AAT	GTT	ACA	CGG	816
Gln	Val	Ala	Val	Cys	Gly	Ser	Leu	Ile	Gly	Trp	Arg	Asn	Val	Thr	Аrg	
			240					245					250			
CTG	CTG	GTG	TTT	TCC	ACA	GAT	GCC	GGG	TTT	CAC	TTT	GCT	GGA	GAT	GGG	864
Leu	Leu	Val	Phe	Ser	Thr	Asp	Ala	Gly	Phe	His	Phe	Ala	Gly	Asp	Gly	
		255					260					265				
AAA	CTT	GGT	GGC	ATT	GTT	TTA	CCA	AAT	GAT	GGA	CAA	TGT	CAC	CTG	GAA	912
Lys	Leu	Gly	Gly	Ile	Yal	Leu	Pro	Asn	Asp	Gly	Gln	Cys	His	Leu	Glu	
	270					275					280					
AAT	AAT	ATG	TAC	ACA	ATG	AGC	CAT	TAT	TAT	GAT	TAT	CCT	TCT	ATT	GCT	960
Asn	Asn	Met	Tyr	Thr	Met	Ser	His	Tyr	Tyr	Asp	Tyr	Pro	Ser	Ile	Ala	
285					290					295					300	-
CAC	CTT	GTC	CAG	AAA	CTG	AGT	GAA	AAT	AAT	ATT	CAG	ACA	ATT	TTT	GCA	1008
His	Leu	Val	Gln	Lys	Leu	Ser	Glu	Asn	Asn	Ile	Gln	Thr	Ile	Phe	Ala	
				305					310					315		
GTT	ACT	GAA	GAA	TTT	CAG	CCT	GTT	TAC	AAG	GAG	CTG	AAA	AAC	TTG	ATC	1056
Val	Thr	Glu	Glu	Phe	Gln	Pro	Val	Tyr	Lys	Glu	Leu	Lys	Asn	Leu	Ile	
			320					325					330			
CCT	AAG	TCA	GCÀ	GTA	GGA	ACA	TTA	TCT	GCA	AAT	TCT	AGC	AAT	GT A	TTA	1104
Pro	Lys	Ser	Ala	Val	Gly	Thr	Leu	Ser	Ala	Asn	Ser	Ser	Asn	Yal	He	
		335					340					345				
CAG	TTG	λТС	λΤΤ	GAT	GCA	TAC	λλΤ	TCC	CTT	TCC	TCA	G A A	GTC	ТТК	TTG	1152
Gln	Leu	He	lle	Asp	Ala	Tyr	λsπ	Ser	Leu	Ser	Ser	Glu	Val	He	Leu	
	350					355					360					

GΛΛ	YVC	GGC	AAA	TTG	TCA	CAA	GGA	GΤλ	λСΛ	λΤλ	AGT	TAC	AAA	TCT	TAC	1200
Glu	λsπ	Gly	Lys	Leu	Ser	Glu	Gly	Val	Thr	[le	Ser	Туr	Lys	Ser	Tyr	
365					370					375					380	
TGC	AAG	AAC	GGG	GTG	AAT	GGA	ACA	GGG	GAA	AAT	GGA	AGA	AAA	TGT	TCC	1248
Cys	Lys	Asn	Gly	Val	Asn	Gly	Thr	Gly	Glu	Asn	Gly	Arg	Lys	Cys	Ser	
				385					390					395		
AAT	ATT	TCC	ATT	GGA	GAT	GAG	GTT	CAA	TTT	GAA	ATT	AGC	ATA	ACT	TCA	1296
Asn	Ile	Ser	Ile	Gly	Asp	Glu	Val	Gln	Phe	Glu	[le	Ser	lle	Thr	Ser	
	,		400					405					410			
AAT	AAG	TGT	CCA	AAA	AAG	GAT	TCT	GAC	AGC	TTT	AAA	ATT	AGG	CCT	CTG	1344
Asn	Lys	Cys	Pro	Lys	Lys	Asp	Ser	Asp	Ser	Phe	Lys	Ile	Arg	Pro	Leu	
		415					420					425				
GGC	TTT	ACG	GAG	GAA	GTA	GAG	GTT	ATT	CTT	CAG	TAC	ATC	TGT	GAA	TGT	1392
Gly	Phe	Thr	Glu	Glu	Val	Glu	Val	Ile	Leu	Gln	Tyr	Ile	Cys	Glu	Cys	
	430					435					440					
GAA	TGC	CAA	AGC	GAA	GGC	ATC	CCT	GAA	AGT	CCC	AAG	TGT	CAT	GAA	GGA	1440
Glu	Cys	Gln	Ser	Glu	Gly	Ile	Pro	Glu	Ser	Pro	Lys	Cys	His	Glu	Gly	
445					450					455					460	
AAT	GGG	ACA	TTT	GAG	TGT	GGC	GCG	TGC	AGG	TGC	AAT	GAA	GGG	CGT	GTT	1488
Asn	Gly	Thr	Phe	Glu	Cys	Gly	Ala	Cys	Arg	Cys	Asn	Glu	Gly	Arg	Val	
			• ,	465		,			470					475		
GGT	AGA	CAT	TGT	GAA	TGC	AGC	ACA	GAT	G A A	GTT	AAC	AGT	GAA	GAC	ATG	1536
Gly	Arg	His	Cys	Glu	Cys	Ser	Thr	Asp	Glu	Val	Asn	Ser	Glu	Asp	Met	
			480					485					490			
GAT	GCT	TAC	TGC	AGG	AAA	GAA	AAC	AGT	TCA	GAA	ATC	TGC	AGT	AAC	AAT	1584
Asp	Ala	Tyr	Cys	Arg	Lys	Glu	Asn	Ser	Ser	Glu	lle	Cys	Ser	Asn	· Asn	
		495					500					505				
GGA	GAG	TGC	GTC	TGC	GGA	СЛG	тст	GTT	TGT	λGG	λλG	ЛGG	GAT	AAT	λСΛ	1632
Gly	Glu	Cys	Val	Cys	Gly	Gln	Cys	Val	Cys	λrg	Lys	Arg	λsp	λsn	Thr	
	5 l 0					515					520					

AAT	GΛA	ገ ፐ ፕ	TAT	TCT	GGC	AAA	TTC	TGC	GAG	TGT	GAT	AAT	TTC	AAC	TGT	1680
λsπ	GLu	lle	Tyr	Ser	Gly	Lys	Phe	Cys	GLu	Cys	Аsр	Asn	Phe	λsn	Cys	
525					530					535					540	
GAT	AGA	TCC	AAT	GGC	ATT	ATT	TGT	GGA	GGA	AAT	GGT	GTT	TGC	AAG	TGT	1728
Asp	Arg	Ser	Asn	Gly	Leu	Ile	Cys	Gly	Gly	Asn	Gly	Val	Cys	Lys	Cys	
				545					550					555		
CGT	GTG	TGT	GAG	TGC	AAC	CCC	AAC	TAC	ACT	GGC	AGT	GCA	TGT	GAC	TGT	1776
Arg	Val	Cys	Glu	Cys	Asn	Pro	Asn	Tyr	Thr	Gly	Ser	λla	Cys	Asp	Cys	
			560					565					570			
TCT	TTG	GAT	ACT	AGT	ACT	TGT	GAA	GCC	AGC	AAC	GGA	CAG	ATC.	TGC	AAT	1824
Ser	Leu	Asp	Thr	Ser	Thr	Cys	Glu	Ala	Ser	Asn	Gly	Gln	Ile	Cys	Asn	
		575					580		•			585				
GGC	CGG	GGC	ATC	TGC	GAG	TGT	GGT	GTC	TGT	AAG	TGT	ACA	GAT	CCG	AAG	1872
Gly	Arg	Gly	Ile	Cys	Glu	Cys	Gly	Yal	Cys	Lys	Cys	Thr	Аsр	Pro	Lys	
	590					595					600					
TTT	CAA	GGG	CAA	ACG	TGT	GAG	ATG	TGT	CAG	ACC	TGC	CTT	GGT	GTC	TGT	1920
Phe	Gln	Gly	G1n	Thr	Cys	Glu	Меt	Cys	Gln	Thr	Cys	Leu	Gly	Yal	Cys	
605					610					615					620	
GCT	GAG	CAT	AAA	GAA	TGT	GTT	CAG	TGC	AGA	GCC	TTC	AAT	AAA	GGA	GAA	1968
Ala	Glu	His	Lys	Glu	Cys	Val	Gln	Cys	Arg	Ala	Phe	Asn	Lys	Gly	Glu	
				625					630					635		
AAG	AAA	GAC	ACA	TGC	ACA	CAG	GAA	TGT	TCC	TAT	TTT	AAC	ATT	ACC	AAG	2016
Lys	Lys	Asp	Thr	Cys	Thr	GIn	Glu	Cys	Ser	Tyr	Phe	Asn	Ile	Thr	Lys	
			640					645					650			
GTA	GAA	AGT	CGG	GAC	AAA	TTA	ccc	CAG	CCG	GTC	CAA	CCT	GAT	CCT	GTG	2064
Yal	Glu	Ser	۸rg	λsp	Lys	Leu	Pro	Gln	Pro	Val	Gln	Pro	Asp	Pro	Val	
		655					660					665				
TCC	CAT	TGT	λλG	GAG	AAG	GAT	GTT	GAC	GAC	TGT	TGG	TTC	TAT	τττ	ACG	2112
Ser	llis	Cys	Lys	Glu	Lys	λsp	Val	λsρ	λsp	Cys	Trp	Phe	Туг	Phe	Thr	
	670					675					680					

TAT TCA GTG AAT GGG AAC AAC GAG GTC ATG GTT CAT GTT GTG GAG AAT	2160
Tyr Ser Val Asn Gly Asn Asn Glu Val Met Val His Val Val Glu Asn	
685 690 695 700	
CCA GAG TGT CCC ACT GGT CCA GAG GAT CCC GAG CTGCTGGAAG CAGGCTCA	GC 2213
Pro Glu Cys Pro Thr Gly Pro Glu Asp Pro Glu	
705 . 710	
GCTCCTGCCT GGACGCATCC CGGCTATGCA GCCCCAGTCC AGGGCAGCAA GGCAGGCC	CC 2273
GTCTGCCTCT TCACCCGGAG CCTCTGCCCG CCCCACTCAT GCTCAGGGAG AGGGTCTT	CT 2333
GGCTTTTTCC CAGGCTCTGG GCAGGCACAG GCTAGGTGCC CCTAACCCAG GCCCTGCA	CA 2393
CAAAGGGGCA GGTGCTGGGC TCAGACCTGC CAAGAGCCAT ATCCGGGAGG ACCCTGCC	CC 2453
TGACCTAAGC CCACCCCAAA GGCCAAACTC TCCACTCCCT CAGCTCGGAC ACCTTCTC	TC 2513
CTCCCAGATT CCAGTAACTC CCAATCTTCT CTCTGCA GAG CCC AAA TCT TGT GA	.C 2568
Glu Pro Lys Ser Cys As	p
715	
AAA ACT CAC ACA TGC CCA CCG TGC CCA GGTAAGCCAG CCCAGGCCTC	2615
Lys Thr His Thr Cys Pro Pro Cys Pro	
720 725	
GCCCTCCAGC TCAAGGCGGG ACAGGTGCCC TAGAGTAGCC TGCATCCAGG GACAGGCC	CCC 2675
AGCCGGGTGC TGACACGTCC ACCTCCATCT CTTCCTCA GCA CCT GAA CTC CTG	2728
Ala Pro Glu Leu Leu	
730	
GGG GGA CCG TCA GTC TTC CTC TTC CCC CCA AAA CCC AAG GAC ACC CTC	2776
Gly Gly Pro Ser Val Phe Leu Phe Pro Pro Lys Pro Lys Asp Thr Le	1
735 740 745	
ATG ATC TCC CGG ACC CCT GAG GTC ACA TGC GTG GTG GAC GTG AG	C 2824
Met Ile Ser Arg Thr Pro Glu Val Thr Cys Val Val Val Asp Val Se	r
750 755 760	
CAC GAA GAC CCT GAG GTC AAG TTC AAC TGG TAC GTG GAC GGC GTG GA	G 2872
His Glu Asp Pro Glu Val Lys Phe Asn Trp Tyr Val Asp Gly Val Gl	
765 770 775	

GTG	CAT	AAT	GCC	λλG	ACA	AAG	CCG	CGG	GAG	GAG	CVC	TAC	AAC	AGC	ACG	2920
Val	His	Asn	Ala	Lys	Thr	Lys	Pro	Arg	Glu	Glu	Gln	Туr	Аsп	Ser	Thr	
780					785					790					795	
TAC	CGG	GTG	GTC	AGC	GTC	CTC	ACC	GTC	CTG	CAC	CAG	GAC	TGG	CTG	AAT	2968
Tyr	Arg	Val	Yal	Ser	Val	Leu	Thr	Val	Leu	His	Gln	Asp	Trp	Leu	Asn	
				800					805					810		
GGC	AAG	GAG	TAC	AAG	TGC	AAG	GTC	TCC	AAC	A A A	GCC	CTC	CCY	GCC	CCC	3016
Gly	Lys	Glu	Tyr	Lys	Cys	Lys	Val	Ser	Asn	Lys	Ala	Leu	Pro	Ala	Pro	
			815					820					825			
ATC	GAG	AAA	ACC	ATC	TCC	AAA	GCC	AAA	GGT	GGGA	ccc (	GTGG(	GTG	CG		3063
Ile	Glu	Lys	Thr	Ile	Ser	Lys	Ala	Lys								
		830					835									
AGG(	GCCAG	CAT (	GGAC	A G A G (	GC C	GGCT	CGGCC	CAC	CCCT	CTGC	CCT	GAGA(	GTG A	CCG	CTGTAC	3123
CAAG	CCTC	rgt (	CTAC	CA GO	GG C	G CC	cc cc	GA GA	AA CO	CA CA	AG G1	G T	AC AC	CC C1	r G	3172
				G	ly Gl	ln Pi	o Ar	rg Gl	lu Pi	co GI	ln Va	al Ty	r Th	ır Le	eu	
							84	10				8 4	15			
CCC	CCA	TCC	CGG	GAT	GAG	CTG	ACC	AAG	AAC	CAG	GTC	AGC	CTG	ACC	TGC	3220
Pro	Pro	Ser	Arg	Asp	Glu	Leu	Thr	Lys	Asn	Gln	Yal	Ser	Leu	Thr	Cys	
		850					855					860				
CTG	GTC	AAA	GGC	TTC	TAT	CCC	AGC	GAC	ATC	GCC	GTG	GAG	TGG	GAG	AGC	3268
Leu	Yal	Lys	Gly	Phe	Tyr	Pro	Ser	Asp	Ile	Ala	Yal	Glu	Trp	Glu	Ser	
	865					870	_				875					
AAT	GGG	CAG	CCG	GAG	AAC	AAC	TAC	AAG	ACC	ACG	CCT	CCC	GTG	CTG	GAT	3316
Asn	Gly	Gln	Pro	Glu	Asn	Asn	Tyr	Lys	Thr	Thr	Pro	Pro	Val	Leu	Asp	
880					885					890					895	
TCC	GAC	GGC	TCC	TTC	TTC	CTC	TAC	AGC	AAG	CTC	ACC	GTG	GAC	ΛAG	AGC	3364
Ser	λsp	Gly	Ser	Phe	Phe	Leu	Туr	Ser	Lys	Leu	Thr	Val	Asp	Lys	Ser	
				900					905					910		

AGG TGG CAG CAG GGG AAC GTC TTC TCA TGC TCC GTG ATG CAT GAG GCT 3412

Arg Trp Gln Gln Gly Asn Val Phe Ser Cys Ser Val Met His Glu Ala
915 920 925

CTG CAC AAC CAC TAC ACG CAG AAG AGC CTC TCC CTG TCT CCG GGT AAA 3460

Leu His Asn His Tyr Thr Gln Lys Ser Leu Ser Leu Ser Pro Gly Lys
930 935 940

Sequence No. 3

Length of sequence: 13

Type of sequence: Amino acid

Topology: Linear

Kind of sequence: Peptide

Sequence

Cys Leu His Gly Pro Glu Ile Leu Asp Val Pro Ser Thr

1 5 10

Sequence No. 4

Length of sequence: 31

Type of sequence: Nucleic acid

Number of strands: Single

Topology: Linear

Kind of sequence: Other nucleic acid, synthetic DNA

Sequence

GCGGATCCCG AGCTGCTGGA AGCAGGCTCA G

31

Sequence No. 5

Length of sequence: 27

Type of sequence: Nucleic acid

Number of strands: Single

Topology: Linear

Kind of sequence: Other nucleic acid, synthetic DNA

Sequence

CCTCTAGACG GCCGTCGCAC TCATTTA

27

Sequence No. 6

Length of sequence: 73

Type of sequence: Nucleic acid

Number of strands: Single

Topology: Linear

Kind of sequence: Other nucleic acid, synthetic DNA

Sequence

CTAGACCACC ATGTTCCCCA CCGAGAGCGC ATGGCTTGGG AAGCGAGGCG CGAACCCGGG

CCCCGGAGCT GCA

73

Sequence No. 7

Length of sequence: 65

Type of sequence: Nucleic acid

Number of strands: Single

Topology: Linear

Kind of sequence: Other nucleic acid, synthetic DNA

Sequence

GCTTCGGGGC CCGGGTTCGC GCCTCGCTTC CCAAGCCATG CGCTCTCGGT GGGGAACATG

GTGGT

65

Sequence No. 8

Length of sequence: 51

Type of sequence: Nucleic acid

Number of strands: Single

Topology: Linear

Kind of sequence: Other nucleic acid, synthetic DNA Sequence

CTCCGGGAGA CGGTGATGCT GTTGCTGTGC CTGGGGGTCC CGACCGGCAG G

51

Sequence No. 9

Length of sequence: 55

Type of sequence: Nucleic acid

Number of strands: Single

Topology: Straight chain

Kind of sequence: Other nucleic acid, synthetic DNA

Sequence

CCTGCCGGTC GGGACCCCCA GGCACAGCAA CAGCATCACC GTCTCCCGGA GTCGA

55

Sequence No. 10

Length of sequence: 37

Type of sequence: Nucleic acid

Number of strands: Single

Topology: Linear

Kind of sequence: Other nucleic acid, synthetic DNA

Sequence

CACTGCAGGC AGGCCTTACA ACGTGGACAC TGAGAGC

37

Sequence No. 11

Length of sequence: 22

Type of sequence: Nucleic acid

Number of strands: Single

Topology: Linear

Kind of sequence: Other nucleic acid, synthetic DNA

Sequence

GCAGAAACCT GTAAATCAGC AG

Sequence No. 12

Length of sequence: 22

Type of sequence: Nucleic acid

Number of strands: Single

Topology: Linear

Kind of sequence: Other nucleic acid, synthetic DNA

Sequence

GCATTTATGC GGAAAGATGT GC

22

Sequence No. 13

Length of sequence: 29

Type of sequence: Nucleic acid

Number of strands: Single

Topology: Linear

Kind of sequence: Other nucleic acid, synthetic DNA

Sequence

CGGGATCCGT GAAATAACGT TTGGGTCTT

29

Sequence No. 14

Length of sequence: 22

Type of sequence: Nucleic acid

Number of strands: Single

Topology: Linear

Kind of sequence: Other nucleic acid, synthetic DNA

Sequence

GCGGAAAAGA TGAATTTACA AC

22

Sequence No. 15

Length of sequence: 27

Type of sequence: Nucleic acid

Number of strands: Single

Topology: Linear

Kind of sequence: Other nucleic acid, synthetic DNA

Sequence

GTGGGATCCT CTGGACCAGT GGGACAC

27

Sequence No. 16

Length of sequence: 10

Type of sequence: Amino acid

Topology: Linear

Kind of sequence: Peptide

Sequence

Gly Pro Glu Ile Leu Asp Val Pro Ser Thr

1 5 10

Sequence No. 17

Length of sequence: 10

Type of sequence: Amino acid

Topology: Linear

Kind of sequence: Peptide

Sequence

1

Gly Pro Glu Ile Leu Glu Val Pro Ser Thr

5

Sequence No. 18

Length of sequence: 6

Type of sequence: Amino acid

Topology: Linear

Kind of sequence: Peptide

## Sequence

Gly Arg Gly Asp Ser Pro 1

5

Sequence No. 19

Length of sequence: 4675

Type of sequence: Nucleic acid

Sequence

ATG	GGG	CCY	GAA	CGG	ACA	GGG	GCC	GCG	CCG	CTG	CCG	CTG	CTG	CTG	GTG	48
Met	Gly	Pro	Glu	Arg	Thr	Gly	Ala	Ala	Pro	Leu	Pro	Leu	Leu.	Leu	Val	
				-25					-20					-15		
TTA	GCG	CTC	AGT	CAA	GGC	ATT	TTA	AAT	TGT	TGT	TTG	GCC	TAC	AAT	GTT	96
Leu	Ala	Leu	Ser	Gln	Gly	Ile	Leu	Asn	Cys	Cys	Leu	Ala	Tyr	Asn	Val	
			-10					- 5					1			
GGT	СТС	CCA	GAA	GCA	AAA	ATA	TTT	TCC	GGT	CCT	TCA	AGT	GAA	CAG	TTT	1-14
Gly	Leu	Pro	Glu	Ala	Lys	Ile	Phe	Ser	Gly	Pro	Ser	Ser	Glu	Gln	Phe	
	5					10					15					
GGG	TAT	GCA	GTG	CAG	CAG	TTT	ATA	AAT	CCA	AAA	GGC	AAC	TGG	TTA	CTG	192
Gly	Tyr	Ala	Val	Gln	Gln	Phe	Ile	Asn	Pro	Lys	Gly	Asn	Trp	Leu	Leu	
20					25					. 30					35	
GTT	GGT	TCA	ССС	TGG	AGT	GGC	TTT	CCT	GAG	AAC	CGA	ATG	GGA	GAT	GTG	240
Val	Gly	Ser	Pro	Trp	Ser	Gly	Phe	Pro	Glu	Asn	Arg	Met	Gly	Asp	Val	
				40					45					50		
TAT	A A A	TGT	CCT	GTT	GAC	CTA	TCC	ACT	GCC	ACA	TGT	GAA	AAA	CTA	AAT	288
Tyr	Lys	Cys	Pro	Val	qsk	Leu	Ser	.Thr	Ala	Thr	Cys	Glu	Lys	Leu	Asn	
			55					60					65			
TTG	СЛА	ACT	ТСЛ	ACA	AGC	λΤΤ	CCY	AAT	GTT	АСТ	GAG	ATG	444	ACC	A A C	336
Leu	Gln	Thr	Ser	Thr	Ser	lle	Pro	λsn	Yal	Thr	Glu	Met	Ĺys	Thr	Λsn	
		70					75					80				

ATG	AGC	CTC	GGC	TTG	ATC	CTC	АСС	AGG	AAC	АTС	GGY	ЛСT	GGA	GGT	TTT	384
Мet	Ser	Leu	Gly	Leu	lle	Leu	Thr	Arg	λsn	Меt	Gly	Thr	Gly	G 1 y	Phe	
	85					90					95					
CTC	ACA	TGT	GGT	ССТ	CTG	TGG	GCA	CAG	CAA	TGT	GGG	TAK	CAG	TAT	TAC	432
Leu	Thr	Cys	Gly	Pro	Leu	Trp	Ala	Gln	Gln	Cys	Gly	Asn	Gln	Туr	Tyr	
100					105					110					-115	
ACA	ACG	GGT	GTG	TGT	TCT	GAC	ATC	AGT	CCT	GAT	TTT	CAG	CTC	TCA	GCC	480
Thr	Thr	Gly	Val	Cys	Ser	Asp	Ile	Ser	Pro	Asp	Phe	Gln	Leu	Ser	Ala	
			٠.	120					125					130		
AGC	TTC	TCA	CCT	GCA	ACT	CAG	ccc	TGC	CCT	TCC	CTC	ATA	GAT	GTT	GTG	528
Ser	Phe	Ser	Pro	Ala	Thr	Gln	Pro	Cys	Pro	Ser	Leu	Ile	Asp	Val	Val	
			135					140					145			
GTT	GTG	TGT	GAT	GAA	TCA	AAT	AGT	ATT	TAT	CCT	TGG	GAT	GCA	GTA	AAG	576
Val	Val	Cys	Asp	Glu	Ser	Asn	Ser	Ile	Tyr	Pro	Trp	Asp	Ala	Val	Lys	
		150					155					160				
AAT	TTT	TTG	GAA	AAA	TTT	GTA	CAA	GGC	CTT	GAT	ATA	GGC	CCC	ACA	AAG	624
Asn	Phe	Leu	Glu	Lys	Phe	Va1	Gln	Gly	Leu	Asp	Ile	Gly	Pro	Thr	Lys	
	165					170			•		175					
ACA	CAG	GTG	GGG	ATT	ATT	CAG	TAT	GCC	AAT	AAT	CCA	AGA	GTT	GTG	TTT	672
Thr	G1n	Val	Gly	Leu	Ile	Gln	Tyr	Ala	Asn	Asn	Pro	Arg	Val	Val	Phe	
180					185					190					195	
AAC	TTG	AAC	ACA	TAT	AAA	ACC	AAA	G A A	GAA	ATG	ATT	GTA	GCA	ACA	TCC	720
Asn	Leu	Asn	Thr	Tyr	Lys	Thr	Lys	Glu	Glu	Меt	Ile	Val	Ala	Thr	Ser	÷
				200					205					210		
CAG	ACA	TCC	CAA	TAT	GGT	GGG	GAC	CTC	ACA	AAC	ACA	TTC	GGA	GCA	ATT	768
Gln	Thr	Ser	Gln	Туг	Gly	Gly	Asp	Leu	Thr	Asn	Thr	Phe	Gly	Ala	lle	
			215					220					225			
CIVIV	TAT	GCA	A G A	AAA	TAT	GCC	TAT	TCA	GCA	GCT	TCT	GGT	GGG	CGA	CGA	816
Gln	Tyr	λla	Arg	Lys	Туг	λla	Туr	Ser	Лlа	Лlа	Ser	Gly	Gly	λrg	λrg	
		230					235				_	240				

AGT	GCT	ACG	AAA	GTA	ATG	GTA	GTT	GTA	ACT	GAC	GGT	$G \Lambda \Lambda$	TCA	CAT	GAT	86-
Ser	Λĺa	Thr	Lys	Val	Met	Val	Val	Val	Thr	Asp	Gly	Glu	Ser	His	λsp	
	245					250					255					
GGT	TCA	ATG	TTG	AAA	GCT	GTG	ATT	GAT	CAA	TGC	AAC	CAT	GAC	AAT	ATA	912
Gly	Ser	Met	Leu	Lys	Ala	Val	lle	Asp	Gln	Cys	Asn	His	dsp	Asn	Ile	
260					265				,	270					.275	
CTG	AGG	TTT	GGC	ATA	GCA	GTT	CTT	GGG	TAC	TTA	AAC	AGA	AAC	GCC	CTT	960
Leu	Arg	Phe	Gly	Ile	Ala	Val	Leu	Gly	Tyr	Leu	Аsп	Arg	Asn	Ala	Leu	
				280					285			,		290		
GAT	ACT	AAA	TAA	TTA	ATA	AAA	GAA	ATA	AAA	GCG	ATC	GCT	AGT.	ATT	CCA	1008
Asp	Thr	Lys	Asn	Leu	Ile	Lys	Glu	Ile	Lys	Ala	lle	Ala	Ser	[le	Pro	
			295					300					305			
ACA	GAA	AGA	TAC	TTT	TTC	AAT	GTG	TCT	GAT	GAA	GCA	GCT	CTA	CTA	GAA	1056
Thr	Glu	Arg	Tyr	Phe	Phe	Asn	Val	Ser	Asp	Glu	Ala	Ala	Leu	Leu	Glu	
		310					315					320				
AAG	GCT	GGG	ACA	TTA	GGA	GAA	CAA	ATT	TTC	AGC	ATT	GAA	GGT	ACT	GTT	1104
Lys	Ala	Gly	Thr	Leu	Gly	Glu	Gln	Ile	Phe	Ser	Ile	Ģlu	Gly	Thr	Yal	
	325					330			•		335					
CAA	GGA	GGA	GAC	AAC	TTT	CAG	ATG	GAA	ATG	TCA	CAA	GTG	GGA	TTC	AGT	1152
Gln	Gly	Gly	Asp	Asn	Phe	Gln	Met	Glu	Met	Ser	Gln	Val	Gly	Phe	Ser	
340					345					350					355	
GCA	GAT	TAC	TCT	TCT	CAA	AAT	GAT	ATT	CTG	ATG	CTG	GGT	GCA	GTG	GGA	1200
Ala	Asp	Tyr	Ser	Ser	Gln	Asn	Asp	lle	Leu	Жet	Leu	Gly	Ala	Val	Gly	
				360					365					370		
GCT	TTT	GGC	TGG	AGT	GGG	ACC	ATT	GTC	CAG	AAG	ACA	TCT	CAT	GGC	CAT	1248
Ala	Phe	Gly	Trp	Ser	Gly	Thr	lle	Yal	Gln	Lys	Thr	Ser	His	Gly	His	
			375					380					385			
TTG	ATC	TTT	CCT	A A A	CAA	GCC	TTT	GAC	CAA	ATT	CTG	CAG	GYC	AGA	AAT	1296
Leu	lle	Phe	Pro	Lys	Gln	Лlа	Phe	λsp	Gln	He	Leu	Gln	λsp	Λrg	λsπ	
		390					395					400				

CAC	AGT	TCA	TAT	TTA	GGT	ТЛС	TCT	GTG	GCT	GCA	ATT	TCT	АСТ	G G A	G A A	1344
His	Ser	Ser	Tyr	Leu	Gly	Tyr	Ser	Val	Ala	Ala	lle	Ser	Thr	Gly	Glu	
	405					410					415					
AGC	ACT	CAC	TTT	GTT	GCT	GGT	GCT	CCT	CGG	GCA	TAK	TAT	ACC	GGC	CAG	1392
Ser	Thr	His	Phe	Yal	Ala	Gly	Ala	Pro	Arg	Ala	Asn	Tyr	Thr	Gly	Cln	
420					425					430					435	
ATA	GTG	CTA	TAT	AGT	GTG	AAT	GAG	AAT	GGC	ААТ	ATC	ACG	GTT	ΑTT	CAG	1440
Ile	۷al	Leu	Tyr	Ser	Val	Asn	Glu	Asn	Gly	Asn	lle	Thr	Val	lle	Gln	r
				440			,		445					450		
GCT	CAC	CGA	GGT	GAC	CAG	ATT	GGC	TCC	TAT	TT, $T$	GGT	AGT	GTG	CTG	TGT	1488
Ala	His	Arg	Gly	Asp	Gln	Ile	Gly	Ser	Tyr	Phe	Gly	Ser	Val	Leu	Cys	
			455					460					465			
TCA	GTT	GAT	GTG	GAT	AAA	GAC	ACC	ATT	ACA	GAC	GTG	CTC	TTG	GTA	GGT	1536
Ser	Val	Asp	.Val	Asp	Lys	Asp	Thr	Ile	Thr	Asp	Val	Leu	Leu	Val	Gly	
		470					475					480				
GCA	CCA	ATG	TAC	ATG	AGT	GAC	CTA	AAG	AAA	GAG	GAA	GGA	AGA	GTC	TAC	1584
Ala	Pro	Met	Tyr	Met	Ser	Asp	Leu	Lys	Lys	Glu	Glu	Gly	Arg	Val	Tyr	
	485					490					495					
CTG	TTT	ACT	ATC	AAA	AAG	GGC	ATT	TTG	GGT	CAG	CAC	CAA	TTT	CTT	GAA	1632
Leu	Phe	Thr	Ile	Lys	Lys	Gly	Ile	Leu	Gly	Gln	His	Gln	Phe	Leu	Glu	
500					505					510					515	
GGC	CCC	GAG	GGC	ATT	G A A	AAC	ACT	CGA	TTT	GGT	TCA	GCA	ATT	GCA	GCT .	1680
Gly	Pro	Glu	Gly	Ile	Glu	Asn	Thr	Arg	Phe	Gly	Ser	Ala	lle	Ala	Ala	
				520					525					530		
CTT	TCA	GAC	ATC	A-A C	ATG	GAT	GGC	TTT	AAT	GAT	GTG	ATT	GTT	GGT	TCA	1728
Leu	Ser	Asp	I1e	Asn	Меt	Лsр	Gly	Phe	λsn	Asp	Val	lle	Val	Gly	Ser	
			535					540					545			
CCA	CTA	GΑΛ	ΛΛT	CYC	λλΤ	TCT	GGA	GCT	G T A	TAC	ТТЛ	TAC	ΛΛΤ	GGT	CAT	1776
Pro	Leu	Glu	λsπ	Gln	λsn	Ser	Gly	λla	J is V	Tyr	He	Туr	λsn	Gly	llis	
		550					555					560				

CVC	GGC	ACT	ATC	CGC	<i>А</i> С <i>А</i>	A A G	TAT	TCC	CAG	AAA	ATC	TTG	GGA	TCC	GAT	1824
Gln	Gly	Thr	Ile	Arg	Thr	Lys	Tyr	Ser	Gln	Lys	lle	Leu	Gly	Ser	Asp	
	565					570					575					
GGA	GCC	TTT	AGG	AGC	CAT	CTC	CAG	TAC	TTT	GGG	AGG	TCC	TTC	GAT	GGC	1872
Gly	Ala	Phe	Arg	Ser	His	Leu	Gln	Tyr	Phe	Gly	Arg	Ser	Leu	λsp	Gly	
580					585					590					595	
TAT	GGA	GAT	TTA	AAT	GGG	GAT	TCC	ATC	ACC	GAT	GTG	TCT	TTA	GGT	GCC	1920
Tyr	Gly	Asp	Leu	Asn	Gly	qsk	Ser	Ile	Thr	Asp	Val	Ser	Ile	Gly	Ala	
	,			600					605					610		
TTT	GGA	CAA	GTG	GTT	CAA	CTC	TGG	TCA	CAA	AGT	TTA	GCT	GAT.	GTA	GCT	1968
Phe	Gly	Gln	Val	Val	Gln	Leu	Trp	Ser	Gln	Ser	Ile	Ala	Аsр	Val	Ala	
			615					620					625			
ATA	GAA	GCT	TCA	TTC	A C A	CCA	GAA	AAA	ATC	ACT	TTG	GTC	AAC	AAG	TAK	2016
Ile	Glu	Ala	Ser	Phe	Thr	Pro	Glu	Lys	Ile	Thr	Leu	Val	Asn	Lys	Asn	
		630					635					640				
GCT	CAG	ATA	ATT	CTC	AAA	CTC	TGĈ	TTC	AGT	GCA	AAG	TTC	AGA	CCT	ACT	2064
Ala	Gln	Ile	Ile	Leu	Lys	Leu	Cys	Phe	Ser	Ala	Lys	Phe	Arg	Pro	Thr	
	645					650			•		655					
AAG	CAA	AAC	AAT	CAA	GTG	GCC	ATT	GTA	TAT	AAC	ATC	ACA	CTT	GAT	GCA	2112
Lys	Gln	Asn	Asn	Gln	Yal	Ala	Ile	Yal	Tyr	Asn	Ile	Thr	Leu	Asp	Ala	
660					665					670					675	
GAT	GGA	TTT	TCA	TCC	AGA	GTA	ACC	TCC	AGG	GGG	TTA	TTT	AAA	GAA	AAC	2160
Asp	Gly	Phe	Ser	Ser	Arg	Val	Thr	Ser	Arg	Gly	Leu	Phe	Lys	Glu	Asn	
				680			`		685					690		
AAT	GAA	AGG	TGC	CTG	CAG	AAG	AAT	ATG	GTA	GTA	TAA	CAA	GCA	CAG	AGT	2208
Asn	Glu	λrg	Cys	Leu	Gln	Lys	Asn	Met	Val	٧al	Asn	Gln	Ala	Gln	Ser	
			695					700					705			
TGC	CCC	GAG	CAC	ATC	ATT	TAT	АТА	CAG	GAG	CCC	тст	GAT	GTT	GTC	Y Y C	2256
Cys	Pro	Glu	llis	lle	lle	Туr	lle	Gin	Glu	Pro	Ser	λsp	Val	Val	λsn	
		710					715					720				

TCT	TTG	GAT	TTG	CGT	GTG	GAC	ATC	AGT	CTG	GAA	γуС	CCT	GGC	АСТ	AGC	2304
Ser	Leu	Asp	Leu	Arg	Yal	Asp	lle	Ser	Leu	Glu	Asn	Pro	Gly	Thr	Ser	
	725					730					735					
CCT	GCC	CTT	GAA	GCC	TAT	TCT	GAG	ACT	GCC	AAG	GTC	TTC	AGT	ATT	CCT	2352
Pro	Ala	Leu	Glu	Ala	Туr	Ser	Glu	Thr	Ala	Lys	Val	Phe	Ser	lle	Pro	
740					745					750					755	
TTC	CAC	AAA	GAC	TGT	GGT	GAG	GAT	GGA	CTT	TGC	TTA	TCT	GAT	CTA	GTC	2400
Phe	His	Lys	Asp	Cys	Gly	Glu	Asp	Gly	Leu	Cys	lle	Ser	Asp	Leu	Val	
				760					765					770		
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Sequence No. 22

Length of sequence: 22

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Number of strands: Single

Topology: Linear

Kind of sequence: Other nucleic acid, synthetic DNA

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Length of sequence: 21

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Number of strands: Single

Topology: Linear

Kind of sequence: Other nucleic acid, synthetic DNA

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Length of sequence: 8

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Topology: Circular

Kind of sequence: Peptide

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Sequence No. 25

Length of sequence: 8

Type of sequence: Amino acid

Topology: Circular

Kind of sequence: Peptide

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Sequence No. 26

Length of sequence: 8

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Topology: Circular

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Sequence

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Length of sequence: 8

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Topology: Circular

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Topology: Circular

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Sequence

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Topology: Circular

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Sequence

l

Cys Lys Leu Lys Gly Thr Met Cys

## Claims

- 1. A chimeric protein comprising the  $\alpha$  chain or  $\beta$  chain of an integrin and the heavy chain or light chain of an immunoglobulin.
- 2. A chimeric protein heterodimer complex, characterized in that a chimeric protein stated in claim 1 comprising the  $\alpha$  chain of an integrin and the heavy chain or light chain of an immunoglobulin and a chimeric protein stated in claim 1 comprising the  $\beta$  chain of the integrin and the heavy chain or light chain of the immunoglobulin are associated with each other.
- 3. A chimeric protein heterodimer complex, according to claim
- 2, wherein the chimeric proteins stated in claim 1 are associated with each other in any of the following combinations (1), (2) and (3):
- (1) An  $\alpha$  chain-immunoglobulin heavy chain- $\beta$  chain-immunoglobulin heavy chain chimeric protein heterodimer complex, in which a chimeric protein comprising the  $\alpha$  chain of an integrin and the heavy chain of an immunoglobulin and a chimeric protein comprising the  $\beta$  chain of the integrin and the heavy chain of the immunoglobulin are associated with each other.
- (2) An  $\alpha$  chain immunoglobulin heavy chain  $\beta$  chain immunoglobulin light chain chimeric protein heterodimer complex, in which a chimeric protein comprising the  $\alpha$  chain of

an integrin and the heavy chain of an immunoglobulin and a chimeric protein comprising the  $\beta$  chain of the integrin and the light chain of the immunoglobulin are associated with each other.

- (3) An  $\alpha$  chain-immunoglobulin light chain- $\beta$  chain-immunoglobulin heavy chain chimeric protein heterodimer complex, in which a chimeric protein comprising the  $\alpha$  chain of an integrin and the light chain of an immunoglobulin and a chimeric protein comprising of the  $\beta$  chain of the integrin and the heavy chain of the immunoglobulin are associated with each other.
- 4. A chimeric protein or a chimeric protein heterodimer complex, according to any one of claims 1 through 3, wherein the  $\alpha$  chain of an integrin is  $\alpha 1$ ,  $\alpha 2$ ,  $\alpha 3$ ,  $\alpha 4$ ,  $\alpha 5$ ,  $\alpha 6$ ,  $\alpha 7$ ,  $\alpha 8$ ,  $\alpha 9$ ,  $\alpha v$ ,  $\alpha L$ ,  $\alpha M$ ,  $\alpha X$ ,  $\alpha IIb$  or  $\alpha E$ .
- 5. A chimeric protein or a chimeric protein heterodimer complex, according to any one of claims 1 through 3, wherein the  $\beta$  chain of an integrin is  $\beta$ 1,  $\beta$ 2,  $\beta$ 3,  $\beta$ 4,  $\beta$ 5,  $\beta$ 6,  $\beta$ 7 or  $\beta$ 8.
- 6. A chimeric protein heterodimer complex, according to claim 2 or 3, wherein the  $\alpha$  chain of an integrin is  $\alpha 4$  or  $\alpha 2$  and the  $\beta$  chain is  $\beta 1$ .
- 7. A chimeric protein or a chimeric protein heterodimer complex, according to any one of claims 1 through 3, wherein the chimeric protein comprising the  $\alpha 4$  of an integrin and the

heavy chain of an immunoglobulin is identified as the amino acid sequence of sequence No. 1.

- 8. A chimeric protein or a chimeric protein heterodimer complex, according to any one of claims 1 through 3, wherein the chimeric protein comprising the  $\alpha$ 2 of an integrin and the heavy chain of an immunoglobulin is identified as the amino acid sequence of sequence No. 19.
- 9. A chimeric protein or a chimeric protein heterodimer complex, according to any one of claims 1 through 3, wherein the chimeric protein comprising the  $\beta$ 1 of an integrin and the heavy chain of an immunoglobulin is identified as the amino acid sequence of sequence No. 2.
- 10. A DNA coding for a chimeric protein stated in claim 1.
- 11. A DNA coding for a chimeric protein stated in claim 1, wherein the  $\alpha$  chain of an integrin is  $\alpha$ 1,  $\alpha$ 2,  $\alpha$ 3,  $\alpha$ 4,  $\alpha$ 5,  $\alpha$ 6,  $\alpha$ 7,  $\alpha$ 8,  $\alpha$ 9,  $\alpha$ 9,  $\alpha$ 0,  $\alpha$ 1,  $\alpha$ 1,  $\alpha$ 1,  $\alpha$ 1,  $\alpha$ 1,  $\alpha$ 1.
- 12. A DNA coding for a chimeric protein stated in claim 1, wherein the  $\beta$  chain of an integrin is  $\beta$ 1,  $\beta$ 2,  $\beta$ 3,  $\beta$ 4,  $\beta$ 5,  $\beta$ 6,  $\beta$ 7 or  $\beta$ 8.
- 13. A DNA, according to claim11, which is identified as the nucleotide sequence of sequence No. 1 or 19.
- 14. A DNA, according to claim 12, which is identified as the nucleotide sequence of sequence No. 2.
- 15. A recombinant vector, wherein a DNA stated in claim 10 is functionally linked to an expression control sequence.

- 16. A recombinant vector, wherein a DNA stated in claim 11 is functionally linked to an expression control sequence.
- 17. A recombinant vector, wherein a DNA stated in claim 12 is functionally linked to an expression control sequence.
- 18. A recombinant vector, wherein a DNA stated in claim 13 is functionally linked to an expression control sequence.
- 19. A recombinant vector, wherein the DNA stated in claim 14 is functionally linked to an expression control sequence.
- 20. An animal cell, comprising being transfected simultaneously by a recombinant vector in which a DNA coding for a chimeric protein comprising the  $\alpha$  chain of an integrin and the heavy chain or light chain of an immunoglobulin is functionally linked to an expression control sequence, and a recombinant vector in which a DNA coding for a chimeric protein comprising the  $\beta$  chain of the integrin and the heavy chain or light chain of the immunoglobulin is functionally linked to an expression control sequence.
- 21. An animal cell, according to claim 20, which is transfected simultaneously by the recombinant vectors stated in claims 16 and 17.
- 22. An animal cell, according to claim 20, which is transfected simultaneously by the recombinant vectors stated in claims 18 and 19.
- 23. A method for producing the chimeric protein heterodimer complex stated in claim 2, comprising culturing the animal

cell stated in claim 20.

- 24. A drug, comprising a chimeric protein or chimeric protein heterodimer complex stated in any one of claims 1 through 9.
- 25. A drug composition, comprising a chimeric protein or chimeric protein heterodimer complex stated in any one of claims 1 through 9.
- 26. A platelet substitute, comprising an isolated extracellular matrix receptor as an active ingredient.
- 27. A platelet substitute, according to claim 26, wherein the extracellular matrix receptor is an integrin.
- 28. A platelet substitute, according to claim 27, wherein the  $\alpha$  chain of an integrin is  $\alpha 1, \alpha 2, \alpha 3, \alpha 4, \alpha 5, \alpha 6, \alpha 7, \alpha 8, \alpha 9, \alpha v, <math>\alpha L$ ,  $\alpha M$ ,  $\alpha X$ ,  $\alpha I I b$  or  $\alpha E$ .
- 29. A platelet substitute, according to claim 27, wherein the  $\beta$  chain of an integrin is  $\beta$ 1,  $\beta$ 2,  $\beta$ 3,  $\beta$ 4,  $\beta$ 5,  $\beta$ 6,  $\beta$ 7 or  $\beta$ 8.
- 30. A platelet substitute, according to claim 27, wherein the integrin is integrin  $\alpha 2\beta 1$ .
- 31. A platelet substitute, according to claim 26, wherein the extracellular matrix receptor is a chimeric protein heterodimer complex comprising an extracellular matrix receptor and an immunoglobulin.
- 32. A platelet substitute, according to claim 31, wherein the chimeric protein heterodimer complex is a chimeric protein heterodimer complex comprising an integrin and an immunoglobulin.

- 33. A platelet substitute, according to claim 32, wherein the chimeric protein heterodimer complex is the chimeric protein heterodimer complex stated in claim 2.
- 34. A platelet substitute, according to claim 33, wherein the chimeric protein heterodimer complex is the chimeric protein heterodimer complex stated in claim 6.
- 35. A platelet substitute, according to any one of claims 26 through 34, wherein the extracellular matrix receptor is bound to a carrier when used.
- 36. A platelet substitute, according to any one of claims 26 through 35, which is hemostatic.
- 37. A method for testing the binding between a chimeric protein heterodimer complex stated in any one of claims 2 to 9, and a ligand or cells, comprising the steps of bringing a chimeric protein heterodimer complex comprising an integrin and an immunoglobulin, and a ligand or cells into contact with each other, to prepare a mixture, and measuring the amount of the chimeric protein heterodimer complex bound to the ligand or cells or the amount of the ligand or cells protein heterodimer complex.
- 38. A method for searching for a substance capable of being bound to an integrin, comprising using a chimeric protein heterodimer complex stated in any one of claims 2 though 9.
- 39. A substance capable of being bound to an integrin, obtained by using the method stated in claim 38.

- 40. A method for searching for a substance which inhibits the binding between an integrin and a ligand, comprising using the method stated in claim 37.
- 41. A method, according to claim 40, wherein the ligand is a fibronectin fragment identified as sequence No. 3 or a collagen.
- 42. A protein, peptide or low molecular weight compound which inhibits the binding between an integrin and a ligand, obtained by using the method stated in claim 40 or 41.
- 43. A method for measuring the amount of a ligand of an integrin, comprising using a chimeric protein heterodimer complex stated in any one of claims 2 through 9.
- 44. A method for identifying an extracellular matrix exposed region, comprising using a chimeric protein heterodimer complex stated in any one of claims 2 through 9.

## Abstract

The present invention provides integrin-immunoglobulin chimeric protein heterodimer complexes in which the  $\alpha$  chain and the  $\beta$  chain of an integrin are stably associated. The obtained integrin-immunoglobulin chimeric protein heterodimer complexes can be directly used as medicines, and can also be used for determining the binding between an integrin and a ligand, and searching for a substance capable of being bound to an integrin and a substance inhibiting the binding between an integrin and a ligand. They can also be used as diagnostic reagents.

Furthermore, it has been found that an integrin isolated with a stably associated structure can be bound to an extracellular matrix under physiological conditions and in the presence of plasma components. Thus, it has been found that an integrin or an extracellular matrix receptor can be applied as a platelet substitute.

Sequence Table

<110> Kainoh, Mie

Tanaka, Toshiaki

<120> Chimeric proteins, their heterodimer complexes, and platelet substitutes

<130> 1102-98

<141> 1998-09-29

<150> PCT/JP98/00370

<151> 1997-01-29

<160> 31

<170> Wordperfect 5.1

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Leu	Phe	Phe	Ala	Ser	Phe	Ser	Leu	Pro	Vai	830	_		ÇIM		83!	
820					825		_	4-4	ككب			ten	gta	ggc		
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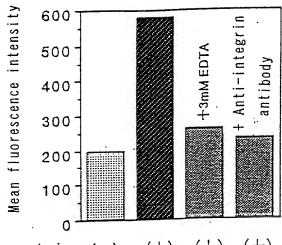
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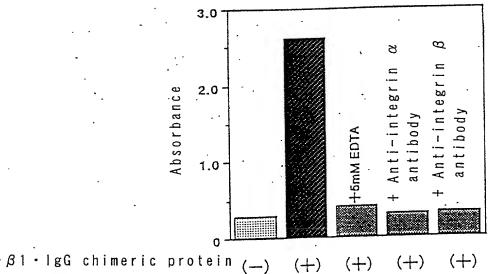
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Fig. 1



(十)  $\alpha$ 4 · IgG- $\beta$ 1 · IgG chimeric protein (—) (十) (十) heterodimer complex

Fig. 2



 $\alpha 4 \cdot \lg G - \beta 1 \cdot \lg G$  chimeric protein (-) (+) heterodimer complex

Fig. 3

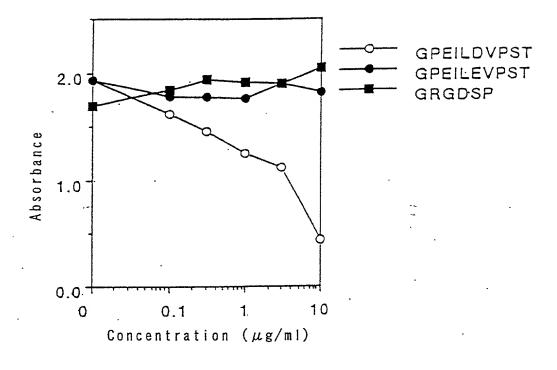


Fig. 4

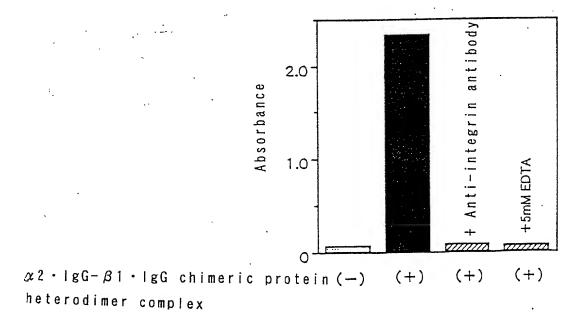
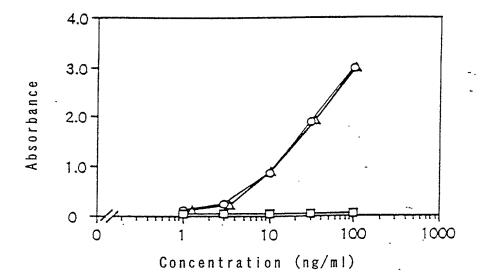


Fig. 5

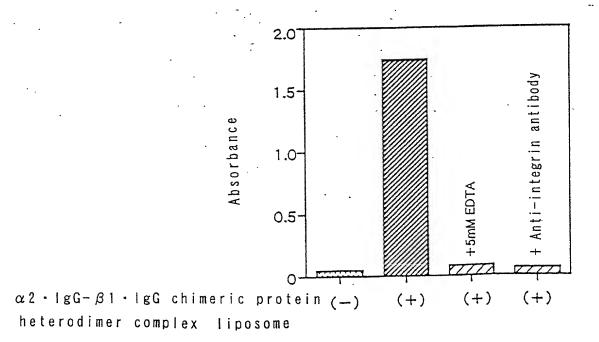


Human IgG liposome (normal plasma)

lntegrin  $\alpha$ 2 $\beta$ 1IgG liposome (normal plasma)

Integrin  $\alpha$ 2 $\beta$ 1IgG liposome (von Willebrand's factor deficient plasma)

Fig. 6



	•
	Original Application
	PCT National Application U.S. Designated Office
	Continuation or Divisional Application
	Continuation-in-Part Application
	COMBINED DECLARATION, POWER OF ATTORNEY AND PETITION
As a be	elow named inventor, I hereby declare that:
My res	idence, post office address and citizenship are as stated below next to my name,
invento the in	Trutes  Trutes
□ wl	nich is described in the specification and claims
	attached hereto.
	filed on
	Application Serial No.
	and was amended on
	(if applicable)
w w	nich is described in International Application No. PCT/JP98/00370
filed	and as amended on
	(if any)

which I have reviewed and for which I solicit a United States patent.

I hereby state that I have reviewed and understand the contents of the above-identified specification, including the claims, as amended by any amendment referred to above.

I acknowledge the duty to disclose information which is material to the examination of this application in accordance with Title 37, Code of Federal Regulations, §1.56(a).

I do not know and do not believe that this invention was ever known or used in the United States before my or our invention thereof or patented or described in any printed publication in any country before my or our invention thereof or more than one year prior to this application or said international application, or in public use or on sale in the United States of America more than one year prior to this application or said international application, or that the invention has been patented or made the subject of an inventor's certificate issued before the date of this application or said international application in any country foreign to the United States of America on an application filed by me or my legal representatives or assigns more than twelve months prior to this application or said international application, or that any application for patent or inventor's certificate on this invention has been filed in any country. Foreign to the United States of America prior to this application or said international application by me or my legal representatives or assigns except as identified below.

## COMBINED DECLARATION, POWER OF ATTORNEY AND PETITION (Page 2)

Attorney Docket No. 1102-98

I hereby claim foreign priority benefits under Title 35, United States Code, §119 of any foreign application(s) for patent or inventor's certificate listed below and have also identified below any foreign applications for patent or inventor's certificate having a filing date before that of the application on which priority is claimed:

Prior Foreign Application(s)

Number	Country	Date of Filing (day,month,year)	Priority Claimed
9-15118	Japan	29 Jan. 1997	■ yes □ no
9-234544	Japan	29 Aug. 1997	■ yes □ no
7 23 13 11			□ yes □ no
			☐ yes ☐ no
			□ yes □ no

application(s) listed below and, is in the prior United States applicate the duty to	nsofar as the subject matter of each cion(s) in the manner provided by the disclose material information as def	of \$120 (as applicable) of any Officer States of the claims of this application is not disclosed first paragraph of Title 35, United States Code, fined in Title 37, Code of Federal Regulations, on(s) and the national or PCT international filing
(Application Serial No.)	(Filing Date)	(Status) (patented, pending, abandoned,
(Application Serial No.)	(Filing Date)	(Status) (patented, pending, abandoned

POWER OF ATTORNEY: As a named inventor, I hereby appoint the following attorneys to prosecute this application and transact all business in the United States Patent and Trademark Office connected therewith:

Austin R. Miller Reg. No. 16,602
T. Daniel Christenbury Reg. No. 31,750
Frank A. Cona Reg. No. 38,412
David A. Sasso Reg. No. 43,084
Patrick J. Farley Reg. No. 42,524

36th Floor 1600 Market Street Philadelphia, PA 19103	DIRECT TELEPHONE CALLS TO ATTORNEY INDICATED ON PAPER: Austin R. Miller T. Daniel Christenbury Frank A. Cona David A. Sasso Patrick J. Farley (215) 563-1810
--	--

## COMBINED DECLARATION, POWER OF ATTORNEY AND PETITION (Page 3)

Attorney Docket No. 1102-98

I hereby petition for grant of a United States Letters Patent on this invention.

I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code and that such willful false statements may jeopardize the validity of the application or any patent issued thereon.

1	1. FULL NAME OF SOLE OR FIRST INVENTOR	INVENTO	or's SIGNATURE	DATE	iv. 4 1998			
1-W	Mie Kainoh RESIDENCE	CITIZENSHIP						
	Kanagawa, Japan	Japan						
	POST OFFICE ADDRESS 633-1-201 Fujisawa, Fujisawa-shi, Kanagawa 251 Japan							
\ \ \ \	DATE				200			
L(X)	Toshiaki Tanaka	Tostial Claude Nov. 4, 1998						
```	RESIDENCE Kanagawa, Japan	citizenship Japan						
<u> </u>	POST OFFICE ADDRESS 11-24 Numama 1-chome, Zushi-shi, Kanagawa 249 Japan							
tu.	3 FULL NAME OF ADDITIONAL JOINT INVENTOR, IF ANY		INVENTOR'S SIGNATURE		DATE			
	RESIDENCE CITIZENSHIP							
<u>.</u>	POST OFFICE ADDRESS							
	4. FULL NAME OF ADDITIONAL JOINT INVENTOR, IF ANY		INVENTOR'S SIGNATURE		DATE			
	RESIDENCE	CITIZE	TIZENSHIP					
	POST OFFICE ADDRESS							
man day man day man day man day man day man day	5. FULL NAME OF ADDITIONAL JOINT INVENTOR, IF ANY		INVENTOR'S SIGNATURE		DATE			
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	6. FULL NAME OF ADDITIONAL JOINT INVENTOR, IF ANY		INVENTOR'S SIGNATURE		DATE			
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	POST OFFICE ADDRESS							
	7. FULL NAME OF ADDITIONAL JOINT INVENTOR, IF ANY		INVENTOR'S SIGNATURE		DATE			
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